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This is a request for filing a PROVISIONAL APPLICATION FOR PATENT under 37 CFR 1.53(c).

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INVENTOR(S)					
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<input checked="" type="checkbox"/> Additional Inventors are being named on the <u>1</u> separately numbered sheets attached hereto					
TITLE OF THE INVENTION (280 characters max)					
SYSTEM AND METHOD FOR MOBILE TICKETING OPERATIONS					
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<input checked="" type="checkbox"/> Specification Number of Pages		65		<input type="checkbox"/> CD(s), Number	
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<input type="checkbox"/> Application Data Sheet. See 37 CFR 1.76					
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Respectfully submitted,

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Date 04 / 15 / 02

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3110 0017

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April 15, 2002

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Assistant Commissioner for Patents
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Crystal Plaza Building 2, Room
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U.S.A.

Dear Sir:

Re: New United States Provisional Patent Application
Title: TB VACCINES
Inventors: LIU, J., CHEN, J., and ALEXANDER, D.C.

We apply in the name of J. Liu, J. Chen, and D.C. Alexander for a provisional patent application entitled **TB VACCINES**.

In addition to the \$150.00 filing fee, included in our firm cheque, we enclose the following documents:

1. provisional application cover page; and
2. patent application

Please direct any questions to Kathryn Schubert at 416-941-9027.

Yours very truly,


Gervas W. Wall
Registration No. 35766

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2002-04-15

Tuberculosis Vaccines Including Recombinant BCG Strains Expressing Alanine Dehydrogenase, Serine Dehydratase and/or Glutamine Synthetase

Field of the Invention

This invention relates to tuberculosis (TB) vaccines.

Background of the Invention

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TB is a deadly contagious disease caused by the infectious agent, *Mycobacterium tuberculosis*. It kills 2 million people each year. The World Health Organization (WHO) 2001 annual report estimated that there would be 8.4 million new TB cases in 1999, up from 8.0 million in 1997. If the present trend continues, it is estimated that between 2000 and 2020, nearly one billion people will be newly infected, 200 million people will become ill and 35 million will die from TB. The spread of HIV/AIDS and the emergence of multidrug-resistant TB contribute to the worsening impact of this disease. Bacille Calmette-Guérin (BCG), an attenuated strain of *Mycobacterium bovis*, is currently the only available vaccine for the prevention of TB. In animal models of infection, BCG vaccination has been demonstrated to induce protective immunity against a *M. tuberculosis* challenge (Baldwins et al., 1998). In humans, BCG vaccination has demonstrated consistent protection against the childhood forms of TB, especially meningitis. However, BCG vaccination is controversial due to variations in its efficacy for protecting adults from pulmonary TB (Fine, 1989; Colditz et al., 1994; Sterne et al., 1998). Trials conducted in the 1940s and 1950s in developed countries such as the United Kingdom, Denmark and North America demonstrated the vaccine to be highly efficient (70-80%). However, in the single largest clinical trial, which took place in India in 1970s and involved more than 265,000 persons, BCG vaccination provided no detectable protection against pulmonary TB. Thus, there is an urgent need to generate an improved vaccine(s) to replace the BCG and to prevent TB.

Several explanations have been suggested for the variation in protective efficacy of BCG (Andersen, 2001). The most prominent hypothesis is that exposure to environmental mycobacteria sensitizes the host against mycobacteria in general, thereby providing

heterologus immunity that obscures the potential benefits of BCG vaccination (Fine, 1995; Fine and Vynnycky, 1998). Furthermore, a recent study showed that the multiplication of BCG was inhibited in animals sensitized with environmental mycobacteria, and consequently BCG vaccination elicited only a transient immune response and failed to provide protective immunity against TB (Brandt et al., 2002). This study also supports the long-standing observation that the induction of immunity to TB requires productive infection by BCG. BCG is a live vaccine; killed BCG does not provide protection. Like *M. tuberculosis*, BCG is capable of forming granulomas and abscesses in various tissues in the infected host (Hogan et al., 2001). The ability of *M. tuberculosis* and *M. bovis* BCG to survive and persist within granulomas, a hostile environment with restricted access to nutrients and reduced oxygen tension, appears to be dependent on the ability of the bacteria to adapt their metabolism to the available source of carbohydrate, nitrogen, and energy (Barclay and Wheeler, 1989). A recent study revealed that fatty acids serve as a source of carbohydrates and are required for persistence of *M. tuberculosis* in mice and activated macrophages (McKinney et al., 2000). Following vaccination in immunocompetent individuals, BCG may persist for certain periods before it is eliminated from the host (Dunn and North 1995; Lagranderie et al., 1996; Moisan et al., 2001).

The key to developing a new and effective TB vaccine is to provide long-term protection (Orme, 2001; Young, 2000). Existing BCG vaccines impart protection against the manifestations of TB in children, but their efficacy wanes over a period of 10 to 15 years, presumably because the protective immunity induced by BCG is gradually lost (Orme, 2001). New strategies to developing an improved vaccine have included the use of attenuated mycobacteria, subunit vaccines and DNA vaccines (Andersen, 2001). However, none of these have proved to be more potent than, or even as effective as BCG. Survival and growth of *M. bovis* BCG is necessary for eliciting protective immunity. It has been shown that early treatment of infected mice with isoniazid to inhibit bacillary growth prevents the development of acquired resistance. BCG strains that persist for extended periods within the host are required in order to obtain more effective vaccines. As such, there is a need for novel, recombinant strains of Bacille Calmette-Guérin.

Summary of the Invention

The invention provides vaccines that overcome the limited ability of BCG strains to use naturally occurring amino acids as the nitrogen source for growth. Furthermore, L-alanine, D-alanine, or L-serine inhibits the growth of BCG strains even when ammonium is present. Expressing a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2] in BCG strains relieves the growth inhibition of BCG by alanine. Similarly, expressing a functional L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6] in BCG strains relieves the growth inhibition of BCG by L-serine. The mechanism for such inhibition occurs through blockage of glutamine synthetase. Overexpression of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] in BCG relieves the growth inhibition of BCG by alanine and L-serine. Recombinant BCG strains that express (or overexpress) a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6], and/or glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] survive and persist longer within the host and consequently induce long-term protective immunity. Such persistent recombinant BCG strains provide more effective vaccines for the prevention of TB and other mycobacterial infections.

The present invention relates to recombinant *Mycobacterium bovis* BCG, which express DNA encoding an alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6], and/or a glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14]. We found that, due to the lack of a functional alanine dehydrogenase [SEQ ID NO:3; SEQ ID NO: 4], BCG cannot utilize alanine (L-alanine or D-alanine) as the only nitrogen source for growth. We further found that alanine (L-alanine or D-alanine) inhibits the growth of all BCG vaccine strains. Said inhibition is relieved by expressing a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2] in BCG. Similarly, BCG cannot utilize L-serine as the only nitrogen source for growth and that growth of BCG is inhibited by L-serine. Expressing a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6] in BCG strains relieves the growth inhibition by L-serine.

Alanine (L-alanine or D-alanine) and L-serine inhibits BCG growth likely by blocking the activity of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14]. Overexpression of

glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] in BCG relieves the growth inhibition of BCG by alanine and L-serine. Glutamine synthetase, in conjunction with glutamate synthase, provides glutamine and glutamate, which are essential for biosynthesis of all amino acids, proteins, purines and pyrimidines. Inhibition of glutamine synthetase stops cell growth. Supplying amino acids that can be converted to glutamate such as L-glutamine, L-glutamate, L-aspartate, and L-asparagine can relieve such inhibition. Indeed, our data show that the inhibition of BCG growth by alanine (L-alanine or D-alanine) or L-serine is relieved by supplementing growth medium with L-glutamine, L-glutamate, L-aspartate, or L-asparagine.

Since BCG is a live vaccine, recombinant BCG strains expressing or overexpressing a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO: 2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO: 6], and/or a glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] survive longer within the human host and subsequently induce long-term memory immunity. These recombinant BCG strains provide extremely useful vaccines.

The present invention relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].

The invention also relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].

The invention further relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8],

[SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13] and [SEQ ID NO:14].

In one embodiment, the live recombinant *Mycobacterium bovis*-BCG strain is selected from the group consisting of *Mycobacterium bovis*-BCG-Russia, *Mycobacterium bovis*-BCG-Moreau, *Mycobacterium bovis*-BCG-Japan, *Mycobacterium bovis*-BCG-Sweden, *Mycobacterium bovis*-BCG-Birkhaug, *Mycobacterium bovis*-BCG-Prague, *Mycobacterium bovis*-BCG-Glaxo, *Mycobacterium bovis*-BCG-Denmark, *Mycobacterium bovis*-BCG-Tice, *Mycobacterium bovis*-BCG-Frappier, *Mycobacterium bovis*-BCG-Connaught, *Mycobacterium bovis*-BCG-Phipps, and *Mycobacterium bovis*-BCG-Pasteur.

Another aspect of the invention is a pharmaceutical composition comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].

The invention also relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].

In yet another aspect of the invention there is a pharmaceutical composition comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8], [SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13] and [SEQ ID NO:14].

In a further aspect of the invention there is a vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].

In another aspect of the invention there is a vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].

In yet another aspect of the invention there is a vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8], [SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13] and [SEQ ID NO:14]. In a preferred embodiment the vaccine or immunogenic composition is for the treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis*. In another preferred embodiment the vaccine or immunogenic compositions of the current invention further comprise a pharmaceutically acceptable carrier. In yet another preferred embodiment the vaccine or immunogenic compositions further comprise adjuvants. In a another embodiment the vaccine or immunogenic compositions further comprises immunogenic materials from one or more other pathogens.

Another aspect of this invention relates to a method for treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis* or *Mycobacterium bovis* comprising administering to the mammal a vaccine or immunogenic composition of the instant invention. In one embodiment the mammal is a cow. In another embodiment the mammal is a human. In yet another embodiment the vaccine or immunogenic composition is administered in the presence of an adjuvant.

A further aspect of the invention is a method for the treatment or prophylaxis of a mammal against cancer comprising administering to the mammal a vaccine or immunogenic composition of the current invention. In one embodiment the cancer is bladder cancer. In another embodiment the vaccine or immunogenic composition is administered in the presence of an adjuvant.

The invention also relates to a test kit comprising the live recombinant *Mycobacterium bovis*-BCG strain of the instant invention.

The invention further relates to a media composition for inhibiting the growth of *Mycobacterium bovis*-BCG comprising alanine as the only nitrogen source for growth. In another embodiment serine is the only nitrogen source for growth. In another embodiment, the media compositions of the current invention further comprise a carbon source, iron, magnesium, and SO_4 . In one embodiment the carbon source is selected from the group consisting of glycerol, dextrose, citrate, and glucose.

The current invention relates to a method for inhibiting the growth of *Mycobacterium bovis*-BCG comprising the steps of (a) obtaining a sample comprising *Mycobacterium bovis*-BCG and (b) culturing the sample in a selective media. In one embodiment the selective media comprises alanine as the only nitrogen source. In yet another embodiment the selective media comprises serine as the only nitrogen source.

Another aspect of the invention relates to a method for culturing *Mycobacterium bovis*-BCG comprising the steps of (a) obtaining a sample comprising *Mycobacterium bovis*-BCG and (b)

culturing the sample in differential media. In one embodiment the differential media comprises histidine.

Brief Description of the Drawings

Preferred embodiments of the invention will be described in relation to the drawings in which:

Fig. 1. Cloning of the *ald* gene. First, a 4.5 kb *ScaI* fragment of *M. tuberculosis* genomic DNA containing the *ald* gene [SEQ ID NO:1] was ligated to *Ecl136II*-linearized pUC19 to generate pUC-ALD. Then, mycobacterial plasmid pALD was created by ligating the 1.9 kb *KpnI* fragment containing the *ald* gene [SEQ ID NO:1] to *KpnI*-linearized pMD31.

Fig. 2. Cloning of the *sdaA* gene.

Cloning of *sdaA* [SEQ ID NO:5] was accomplished in two steps. First, a 9.5 kb *BamHI* fragment of *M. tuberculosis* genomic DNA was ligated to *BamHI*-linearized pMD31 to generate pSDA1. Plasmid pSDAA was generated by cleavage of pSDA1 with *PstI*, followed by self-ligation of the 10.9 kb *PstI* fragment.

Fig. 3. Inhibition of BCG growth by L-alanine in GAS. BCG-Japan, BCG-Frappier, and BCG-Pasteur grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were each inoculated into duplicated 5 ml culture volumes of GAS, GAS without L-alanine, and GAS supplemented with 27 mM L-asparagine, to a cell density of 2×10^7 cells/ml. Cultures were incubated at 37°C with constant shaking for 16 days and then 2 ml aliquots of cell culture were centrifuged and cell pellet lyophilized to determine cell dry weight.

Fig. 4. Inhibition of BCG growth by increasing concentrations of L-alanine in Sauton containing NH_4Cl (5 g/liter). a) BCG-Japan, b) BCG-Frappier, and c) BCG-Pasteur, grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media. Cells were washed and resuspended in Sauton basal medium (no nitrogen source).

Resuspended cells of each strain were inoculated into duplicate 5 ml culture volumes of Sauton media supplemented with NH_4Cl and increasing concentrations of L-alanine. Cultures were incubated at 37°C with constant shaking for 30 days and cell dry weight was determined.

Fig. 5. Inhibition of BCG growth by D-alanine in GAS. BCG-Japan, BCG-Frappier, and BCG-Pasteur grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were each inoculated into 5ml culture volumes of GAS in which L-alanine was replaced by D-alanine, GAS without L-alanine and, GAS (containing D-alanine) supplemented with 27 mM L-asparagine, to a cell density of 2×10^7 cells/ml. Cultures were incubated at 37°C with constant shaking for 13 days and cell dry weight was determined.

Fig. 6. Growth of recombinant BCG strains expressing alanine dehydrogenase [SEQ ID NO:1] in GAS medium. The growth of BCG-Frappier/*ald*, BCG-Pasteur/*ald*, BCG-Frappier/pMD31, BCG-Pasteur/pMD31, BCG-Frappier, and BCG-Pasteur were compared. Cells of each strain, grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were washed and resuspended in Sauton basal medium (no nitrogen source). Resuspended cells were inoculated into duplicate 5 ml culture volumes of GAS without L-alanine, GAS containing L-alanine and GAS in which L-alanine was replaced by D-alanine. Cultures were incubated at 37°C with constant shaking for 15 days and cell dry weight was then determined.

Fig. 7. Inhibition of BCG growth by L-serine in GAS. BCG-Japan, BCG-Frappier, and BCG-Pasteur grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were each inoculated into duplicate 5 ml culture volumes of GAS in which L-alanine was replaced by L-serine, GAS without L-alanine, and GAS (containing L-serine) supplemented with 27 mM L-asparagine, to a cell density of 2×10^7 cells/ml. Cultures were incubated at 37°C with constant shaking for 15 days and cell dry weight was then determined.

Fig. 8. Growth of recombinant BCG strains expressing L-serine dehydratase [SEQ ID NO:5] in GAS medium containing L-serine. The growth of BCG-Japan/*sdaA*,

BCG-Frappier/*sdaA*, BCG-Pasteur/*sdaA*, BCG-Japan, BCG-Frappier, and BCG-Pasteur were compared. Cells of each strain, grown to stationary phase in 7H9/ADC/glycerol/Tween-80 liquid media, were washed and resuspended in Sauton basal medium (no nitrogen source). Resuspended cells were inoculated into duplicate 5 ml culture volumes of GAS without L-alanine, GAS in which L-alanine was replaced by L-serine, and GAS (containing L-serine) supplemented with 27 mM L-asparagine. Cultures were incubated at 37°C with constant shaking for 15 days and cell dry weight was then determined.

Fig. 9. Alignment of A) nucleotide and B) amino acid sequences of the *ald* genes of *Mycobacterium tuberculosis* (*M. tb*) [SEQ ID NO:1; SEQ ID NO:2] and *Mycobacterium bovis* (*M. bovis*) [SEQ ID NO:3; SEQ ID NO:4]. The point deletion causing the frameshift mutation in *M. bovis ald* [SEQ ID NO:3] is indicated with an arrow. Nucleotide codons and amino acids affected by this mutation are highlighted.

Detailed Description of the Invention

BCG vaccine strains have a limited ability to utilize amino acids as the nitrogen source for growth. Furthermore, we found that naturally occurring amino acids L-alanine and L-serine inhibit the growth of BCG strains. Expressing a functional L-alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG relieves the growth inhibition by alanine. Expressing of a functional L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6] in BCG relives the growth inhibition by L-serine. As well, overproduction of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] relieves the growth inhibition by alanine and serine. These novel findings are significant because recombinant BCG strains that express (or overexpress) a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6], and/or glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] will survive better within the human host, induce long-term memory immunity and provide for more effective vaccines to prevent TB, particularly for protecting against pulmonary TB in adults.

It has long been known that administration of killed BCG strains results in a weak and transient immune response. Protective immunity requires survival and replication of BCG

in the vaccinated host. This notion is reinforced by a recent study of an animal model of infection, which showed that prior exposure to live environmental mycobacteria blocked the multiplication of BCG in infected mice. Consequently BCG elicited only a transient immune response which failed to provide protective immunity against TB (Brandt et al., 2002). Live BCG continuously secrete many different antigens that are likely important for the induction of protective immunity. The continuous production of numerous antigens by multiplying BCG gives live vaccines an advantage over subunit vaccines or DNA vaccines which transiently produce a few antigens. Thus the ability of BCG to multiply and persist within the host is an important determinant of BCG efficacy.

In order to grow and persist within the host, BCG must be able to utilize the available nutrients inside the host. It was demonstrated that isocitrate lyase, an essential enzyme for catabolism of fatty acids, is required for persistence of *M. tuberculosis* during the chronic phase of infection and that this requirement was dependent on an intact immune response of the host (McKinney et al., 2000). In another study, an *M. bovis* BCG strain lacking anaerobic nitrate reductase, an enzyme essential for nitrate respiration, failed to persist in lungs, liver and kidneys of immune-competent mice (Fritz et al., 2002). Our findings, that BCG strains utilize only a few types of amino acids as the nitrogen source for growth, and that the growth of all BCG strains are inhibited by naturally occurring L-alanine and L-serine, suggest that the ability of BCG to grow and persist within the host is restricted. The concentration of L-alanine that is available to BCG growing in human is estimated to be 0.33-0.42 mM (Barclay and Wheeler, 1989), which is sufficient to inhibit the growth of BCG-Pasteur or BCG-Frappier, and significantly reduce the growth of BCG-Japan (Fig. 4). The concentration of L-serine present in the extracellular fluids of the host is around 0.1 mM (Barclay and Wheeler, 1989), which may cause significant inhibition of BCG growth. Since multiplication of BCG is required to generate protective immunity, such inhibition by amino acids within the host may prevent the development of long-term protective immunity and hence the lack of protection against pulmonary TB in adults.

M. bovis BCG is also used in the treatment of bladder cancer. Numerous randomized controlled clinical trials indicate that intravesical administration of BCG can prevent or delay tumour recurrence (reviewed in Lamm, 2000; Lockyer and Gillatt, 2001). The

details of how BCG exerts this effect remain to be determined. However, the antitumour response requires an intact T-cell response, and involves increased expression of Th1-type cytokines, including TNF α and IL-6 (reviewed in Prescott et al, 2000). The most effective treatment regimes involve multiple applications of BCG, which suggests that prolonged exposure to the bacteria is required. Similarly, tumours that retain the ability to phagocytize BCG are most susceptible to this treatment (de Boer et al 1996), indicating that bacterial interactions with the tumour are important. As such, a BCG strain demonstrating increased persistence may provide enhanced antitumour activity.

We show that the absence of a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] is responsible for the failure of BCG strains to utilize alanine (L-alanine or D-alanine) as the only nitrogen source. A gene (Rv2708) coding for a L-alanine dehydrogenase (*ald*) [SEQ ID NO:1] was identified in the genome of *M. tuberculosis*. The activity of this enzyme from *M. tuberculosis* had been demonstrated biochemically *in vitro*. Ald converts L-alanine to pyruvate and ammonium, and is highly specific for L-alanine (Hutter and Singh, 1999). This enzyme was detected in the culture supernatant fraction of *M. tuberculosis* but not in *M. bovis* BCG-Japan nor BCG-Copenhagen, even though DNA Southern blot showed that the gene is present in both BCG strains (Anderson et al., 1992). Similarly, we do not detect alanine dehydrogenase activity in any of the 12 BCG strains listed in this report (data not shown). This lack of a functional alanine dehydrogenase in BCG strains is probably caused by a mutation within the *ald* gene [SEQ ID NO:3], and probably originated with the original *M. bovis* strain. A frame-shift mutation is found within the *ald* gene in the published genome sequence of *M. bovis* (Fig. 9) [SEQ ID NO:3]. As a result, the full length L-alanine dehydrogenase protein [SEQ ID NO:2; SEQ ID NO:4] cannot be made in BCG strains and subsequently BCG cannot catabolize alanine. Similarly, the failure of BCG to utilize L-serine as the only nitrogen source is likely to be caused by either mutations or altered expression of the *sdaA* gene [SEQ ID NO:5; SEQ ID NO:6], which encodes L-serine dehydratase. Expression of *sdaA* [SEQ ID NO:5; SEQ ID NO:6] of *M. tuberculosis* in BCG allows BCG strains to grow on L-serine as the only nitrogen source and relieves the inhibition of BCG growth by L-serine (Fig. 8). The inhibition of BCG growth by alanine and serine is

caused by inhibition of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14]. Overexpression of a glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] in BCG relieves the growth inhibition by L-serine, L-alanine and D-alanine.

BCG-Frappier and BCG-Pasteur are more susceptible than BCG-Japan to inhibition by alanine, presumably due to difference in the expression level or activity of glutamine synthetase. BCG-Japan differs from BCG-Frappier or BCG-Pasteur genetically (Behr et al., 1999). Calmette and Guérin developed the BCG vaccine in 1921 after 13 years and 230 passages of an isolate of *M. bovis in vitro*. Starting from 1924, BCG lots were distributed to laboratories around the world. These laboratories continued the passage of the bacteria *in vitro* employing a variety of different recipes and protocols until 1961 when lyophilized seeds were established. As a consequence of such practices, different BCG progeny strains were created, which differed biochemically and genetically (Oettinger et al., 1999; Behr et al., 1999). Our data show that the ability of BCG strains to utilize amino acids as nitrogen source vary; for example, BCG-Japan is able to grow on cationic amino acids including L-arginine and L-lysine while BCG-Pasteur and BCG-Frappier cannot. These differences may also contribute to the differences of BCG efficacy in various clinical trials.

In summary, we use recombinant BCG strains that express (or overexpress) a functional alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], a L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6], and/or glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO: 14] as vaccines to prevent TB and other mycobacterial infections. These recombinant BCG vaccines will induce long-term protective immunity against TB.

Variations of Nucleic Acid Molecules

Modifications

Many modifications may be made to the nucleic acid molecule DNA sequences disclosed in this application and these will be apparent to one skilled in the art. The invention includes nucleotide modifications of the sequences disclosed in this application (or fragments thereof) that are capable of directing expression in bacterial or mammalian

cells. Modifications include substitution, insertion or deletion of nucleotides or altering the relative positions or order of nucleotides.

Sequence Identity

The nucleic acid molecules of the invention also include nucleic acid molecules (or a fragment thereof) having at least about: 70% identity, at least 80% identity, at least 90% identity, at least 95% identity, at least 96% identity, at least 97% identity, at least 98% identity or, most preferred, at least 99% or 99.5% identity to a nucleic acid molecule of the invention and which are capable of expression of nucleic acid molecules in bacterial or mammalian cells. Identity refers to the similarity of two nucleotide sequences that are aligned so that the highest order match is obtained. Identity is calculated according to methods known in the art. For example, if a nucleotide sequence (called "Sequence A") has 90% identity to a portion of [SEQ ID NO: 1], then Sequence A will be identical to the referenced portion of [SEQ ID NO: 1] except that Sequence A may include up to 10 point mutations (such as substitutions with other nucleotides) per each 100 nucleotides of the referenced portion of [SEQ ID NO: 1].

Sequence identity (each construct preferably without a coding nucleic acid molecule insert) is preferably set at least about: 70% identity, at least 80% identity, at least 90% identity, at least 95% identity, at least 96% identity, at least 97% identity, at least 98% identity or, most preferred, at least 99% or 99.5% identity to the sequences provided in SEQ ID NO:1 to SEQ ID NO:14 or its complementary sequence). Sequence identity will preferably be calculated with the GCG program from Bioinformatics (University of Wisconsin). Other programs are also available to calculate sequence identity, such as the Clustal W program (preferably using default parameters; Thompson, JD et al., Nucleic Acid Res. 22:4673-4680), BLAST P, BLAST X algorithms.

Hybridization

The invention includes DNA that has a sequence with sufficient identity to a nucleic acid molecule described in this application to hybridize under stringent hybridization conditions (hybridization techniques are well known in the art). The present invention

also includes nucleic acid molecules that hybridize to one or more of the sequences in [SEQ ID NO:1] to [SEQ ID NO:14] or its complementary sequence. Such nucleic acid molecules preferably hybridize under high stringency conditions (see Sambrook et al. Molecular Cloning: A Laboratory Manual, Most Recent Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.). High stringency washes have preferably have low salt (preferably about 0.2% SSC) and a temperature of about 50-65 °C.

Vaccines

One skilled in the art knows the preparation of live recombinant vaccines. Typically, such vaccines are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection may also be prepared. The preparation may also be emulsified, or the protein encapsulated in liposomes. The live immunogenic ingredients are often mixed with excipients that are pharmaceutically acceptable and compatible with the active ingredient. Suitable excipients are, for example, water, saline, dextrose, glycerol, ethanol, or the like and combinations thereof. In addition, if desired, the vaccine may contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, and/or adjuvants that enhance the effectiveness of the vaccine. Examples of adjuvants which may be effective include but are not limited to: aluminum hydroxide, N-acetyl-muramyl-L-threonyl-D-isoglutamine (thr-MDP), N-acetyl-nor-muramyl-L-alanyl-D-isoglutamine (CGP 11637, referred to as nor-MDP), N-acetylmuramyl-L-alanyl-D-isoglutaminyl-L-alanine-2-(1'-2'-dipalmitoyl-sn -glycero-3-hydroxyphosphoryloxy)-ethylamine (CGP 19835A, referred to as MTP-PE), and RIBI, which contains three components extracted from bacteria, monophosphoryl lipid A, trehalose dimycolate and cell wall skeleton (MPL+TDM+CWS) in a 2% squalene/Tween 80™ emulsion.

The effectiveness of an adjuvant may be determined by measuring the amount of antibodies directed against an immunogenic polypeptide containing a *Mycobacterium tuberculosis* antigenic sequence resulting from administration of the live recombinant *Mycobacterium bovis*-BCG vaccines that are also comprised of the various adjuvants. The vaccines are conventionally administered parenterally, by injection, for example, either subcutaneously or intramuscularly. Additional formulations which are suitable for

other modes of administration include suppositories and, in some cases, oral formulations. For suppositories, traditional binders and carriers may include, for example, polyalkylene glycols or triglycerides; such suppositories may be formed from mixtures containing the active ingredient in the range of 0.5% to 10%, preferably 1%-2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, and the like. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain 10%-95% of active ingredient, preferably 25%-70%.

The vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be prophylactically and/or therapeutically effective.

The vaccine may be given in a single dose schedule, or preferably in a multiple dose schedule. A multiple dose schedule is one in which a primary course of vaccination may be with 1-10 separate doses, followed by other doses given at subsequent time intervals required to maintain and or reinforce the immune response, for example, at 1-4 months for a second dose, and if needed, a subsequent dose(s) after several months. The dosage regimen will also, at least in part, be determined by the need of the individual and be dependent upon the judgment of the practitioner.

In addition, the live recombinant *Mycobacterium bovis*-BCG vaccine administered in conjunction with other immunoregulatory agents, for example, immune globulins. A subject of the present invention is also a multivalent vaccine formula comprising, as a mixture or to be mixed, a live recombinant *Mycobacterium bovis*-BCG vaccine as defined above with another vaccine, and in particular another recombinant live recombinant *Mycobacterium bovis*-BCG vaccine as defined above, these vaccines comprising different inserted sequences.

Pharmaceutical compositions

The pharmaceutical compositions of this invention are used for the treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis* or *Mycobacterium bovis*. The pharmaceutical compositions of this invention are also used to treat patients having degenerative diseases, disorders or abnormal physical states such as cancer.

The pharmaceutical compositions can be administered to humans or animals by methods such as tablets, aerosol administration, intratracheal instillation and intravenous injection.

Media Compositions

The media compositions of this invention for inhibiting the growth of *Mycobacterium bovis*-BCG comprise alanine or serine as the only nitrogen source. When alanine is the only nitrogen source it is present in an amount of at least 0.03mM and when serine is the only nitrogen source it is present in an amount of at least 0.03mM.

The media compositions may further contain carbon in an amount of about 1.35g/L to about 1.65g/L, preferably in an amount of at least 1.5g/L; iron in an amount of about 0.045g/L to about 0.055g/L, preferably in an amount of at least 0.05g/L; magnesium in an amount of about 0.45g/L to about 0.55g/L, preferably in an amount of at least 0.5g/L; and SO₄ in an amount of about 0.045g/L to about 0.055g/L, preferably in an amount of at least 0.05g/L.

Kits

Kits suitable for immunodiagnosis and containing the appropriate labeled reagents are constructed by packaging the appropriate materials, including the live recombinant *Mycobacterium bovis*-BCG strains of the instant invention, in suitable containers, along with the remaining reagents and materials required for the conduct of the assay, as well as a suitable set of assay instructions. Any immunological test format is contemplated, such as ELISA, Western blot, sandwich assay etc., which are well known to those skilled in the art.

Materials and Methods

Bacterial strains and culture conditions. Twelve *M. bovis* BCG strains: BCG-Japan, BCG-Russia, BCG-Moreau, BCG-Sweden, BCG-Birkhaug BCG-Frappier, BCG-Pasteur, BCG-Glaxo, BCG-Phipps, BCG-Tice, BCG-Denmark, and BCG-Prague were used in this study and were obtained from Dr. Marcel Behr (McGill University). The identities of these strains were described in detail previously (Behr et al., 1999). Middlebrook 7H9 medium (Difco) contains (per liter) ammonium sulfate, 0.5 g; L-glutamate, 0.5 g; sodium citrate 0.1 g; pyridoxine, 1 mg; biotin, 0.5 mg; disodium phosphate 2.5g; monopotassium phosphate, 1 g; ferric ammonium citrate 40 mg; magnesium sulfate 50 mg; calcium chloride 0.5 mg; zinc sulfate 1 mg; copper sulfate, 1 mg; and glycerol, 2 ml; with 5 g of albumin (fraction V; bovine), 2 g of dextrose, and 0.05% Tween 80 added after sterilization. Sauton medium contains (per liter) L-asparagine, 4 g; monopotassium sulfate, 0.5 g; magnesium sulfate 0.5 g; ferric ammonium citrate 50 mg; citric acid, 2 g; zinc sulfate, 1 mg; and glycerol, 60 ml; with 0.05% Tween 80 added after sterilization. Glycerol-alanine-salts (GAS) medium contains (per liter) 2 g of ammonium chloride, 1 g of L-alanine, 0.3 g of Bacto Casitone (Difco), 4 g of dibasic potassium phosphate, 2 g of citric acid, 50 mg of ferric ammonium citrate, 1.2 g of magnesium chloride hexahydrate, 0.6 g of potassium sulfate, 1.8 ml of 10 M sodium hydroxide, and 10 ml of glycerol. Tween 80 was added to 0.05% after sterilization. BCG cultures were grown at 37°C with constant shaking for 3-4 weeks.

Cloning of *ald*. Cloning of *ald* [SEQ ID NO:1] was accomplished in two steps (Fig. 1). First, a 4.5kb *ScaI* fragment of *M. tuberculosis* genomic DNA containing *ald* was ligated to *Ecl*136II-linearized pUC19 to generate pUC-ALD. Then mycobacterial plasmid pALD was created by ligating the 1.9 kb *KpnI* fragment containing the *ald* gene [SEQ ID NO:1] to *KpnI*- linearized pMD31 (Yu et al., 1998). The plasmid pALD was introduced by electroporation into *M. bovis* BCG, and recombinant *M. bovis* BCG selected on Middlebrook 7H9 agar (Difco) supplemented with 10% oleic/albumin/dextrose/catalase (OADC) enrichment and 25 µg/ml kanamycin.

Cloning of *sdaA*. Cloning of *sdaA* [SEQ ID NO:5] was accomplished in two steps. First, a 9.5 kb *Bam*HI fragment of *M. tuberculosis* genomic DNA was ligated to *Bam*HI-linearized pMD31 to generate pSDA1. Plasmid pSDAA was generated by cleavage of pSDA1 with *Pst*I, followed by self-ligation of the 10.9 kb *Pst*I fragment. The plasmid pSDAA was introduced by electroporation into *M. bovis* BCG, and recombinant *M. bovis* BCG selected on Middlebrook 7H9 agar (Difco) supplemented with 10% oleic/albumin/dextrose/catalase (OADC) enrichment and 25 µg/ml kanamycin.

Example 1

Growth of BCG strains in Glycerol-Alanine-Salts (GAS) medium. During the course of our studies, we found that BCG-Japan strain was able to grow in GAS medium, albeit slower than in 7H9 medium. BCG-Frappier and BCG-Pasteur could not grow in GAS medium, even after prolonged incubation (2 months). The growth of other BCG strains in GAS medium was also examined. The results are summarized in Table I, and show that BCG-Japan, BCG-Russia, BCG-Moreau, BCG-Sweden and BCG-Birkhaug were able to grow in GAS medium while BCG-Frappier, BCG-Pasteur, BCG-Glaxo, BCG-Phipps, BCG-Tice, BCG-Denmark, and BCG-Prague could not. This is an interesting observation since all 12 BCG strains listed above were able to grow in 7H9 and Sauton broth medium (Table I). To find out why certain BCG strains were unable to grow in GAS medium, the chemical compositions of GAS, 7H9 and Sauton medium were compared. Supplementing ZnSO₄ (1 mg/liter), which is present in 7H9 and Sauton but not in GAS medium, or sodium pyruvate (0.5%), which is required for growth of large colonies of *M. bovis*, did not support the growth of BCG strains in GAS (data not shown). Next, nitrogen sources were compared. L-Asparagine (4 g/liter) is the only nitrogen source in Sauton medium while ammonium chloride (2 g/liter) and L-alanine (1 g/liter) are the main nitrogen sources in GAS. When L-asparagine (at 4 g per liter) was added to GAS medium, BCG-Frappier, BCG-Pasteur, BCG-Glaxo, BCG-Phipps, BCG-Tice, BCG-Denmark, and BCG-Prague were able to grow rapidly (Table I). Supplementing L-aspartate, L-glutamine, or L-glutamate but not other types of amino acids to GAS medium also supported the growth of these BCG strains (Table I). These results show that the failure

of certain BCG strains to grow in GAS medium is caused by their inability to utilize the nitrogen source present.

Example 2

Amino acids as the nitrogen source for growth of BCG strains. The above result prompted us to examine the ability of BCG strains to utilize various types of amino acids as the only nitrogen source. Since GAS medium contains a small amount of Bacto Casitone (0.3 g/liter), which is a complex mixture of various amino acids and peptides, we chose Sauton medium, which is a defined medium, for this purpose. The L-asparagine in the original formula for Sauton medium was replaced individually by each type of amino acids at the same concentration (27 mM), and pH was adjusted to 7.0. Ammonium chloride at 27 mM or 1 mM as the only nitrogen source was also tested. Table II summarizes the results for three representative BCG strains, BCG-Japan, BCG-Pasteur, and BCG-Frappier. Consistent with the result in Table I, all three BCG strains grew rapidly when L-asparagine, L-aspartate, L-glutamine, or L-glutamate was used as the only nitrogen source. BCG-Japan was able to grow on cationic amino acids (e.g., L-arginine, L-lysine) while BCG-Pasteur and BCG-Frappier could not. More interestingly, none of the BCG strains were able to utilize L-alanine, L-serine, L-leucine, L-isoleucine, L-methioine, or L-glycine as the only nitrogen source, while other *Mycobacterium* species, including pathogenic *M. tuberculosis* and *M. avium*, and nonpathogenic *M. smegmatis*, were able grow on these amino acids. These results demonstrate that BCG vaccine strains utilize limited types of amino acids as the nitrogen source for growth; some BCG strains such as BCG-Pasteur or BCG-Frappier can grow only on 4 types of amino acids (Table II). Such a limitation is likely to restrict the ability of BCG to grow and persist *in vivo* (within the host).

Example 3

L-Alanine, D-alanine, or L-serine inhibits the growth of BCG. One surprising finding from the above experiment was that all BCG strains are able to grow on ammonium chloride as the only nitrogen source at both low (1 mM) or high concentrations (27 mM) (Table II). This is contradictory to the result obtained in GAS medium, in which

ammonium chloride at 37 mM does not support the growth of BCG-Pasteur and BCG-Frappier (Table I). Since GAS medium also contains L-alanine, and L-alanine is not utilized by BCG strains for growth (Table II), the only possible explanation is that L-alanine actually inhibits the growth of BCG strains. To prove this, a modified GAS medium, in which L-alanine was omitted, was made and the growth of BCG strains in this medium was examined. As predicted, BCG-Frappier and BCG-Pasteur, which are unable to grow in the original GAS medium containing L-alanine, grew rapidly in GAS without L-alanine (Fig. 3). BCG-Japan also grew more rapidly in this L-alanine free medium than in the original GAS medium (Fig. 3). The same results were obtained for the other nine BCG strains listed in this report.

To further confirm this result, increasing concentrations of L-alanine were added to Sauton medium containing ammonium chloride (5 g/liter) and the growth of BCG-Japan, BCG-Frappier and BCG-Pasteur was determined (Fig. 4). Strikingly, even at a very low concentration (0.25 mM), L-alanine completely inhibited the growth of BCG-Frappier and BCG-Pasteur. Although the growth inhibition of BCG-Japan was somewhat less severe, L-alanine at 0.5 mM significantly reduced its growth and at 8-16 mM the growth was completely inhibited (Fig. 4). Taken together, these results clearly demonstrate that L-alanine inhibits the growth of BCG strains. We further found that D-alanine also inhibits the growth of BCG strains. The presence of D-alanine in GAS medium stopped the growth of BCG-Pasteur and BCG-Frappier, and significantly reduced the growth of BCG-Japan (Fig. 5). Similarly, the presence of L-serine in GAS medium significantly inhibited the growth of BCG-Japan, BCG-Frappier, and BCG-Pasteur (Fig. 7).

Example 4

Expressing L-alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG relieves the inhibition of BCG growth by L-alanine and D-alanine. Alanine is an excellent source of nitrogen for many *Mycobacterium* species including *M. tuberculosis*, *M. avium*, and *M. smegmatis*. D-Alanine degradation begins with racemization to L-alanine, which is then broken down to ammonium and pyruvate by L-alanine dehydrogenase. Interestingly, a functional L-alanine dehydrogenase was detected in *M.*

tuberculosis and *M. smegmatis* but not in BCG-Japan or BCG-Copenhagen (Andersen et al., 1992; Hutter and Dick, 1998). We did not detect L-alanine dehydrogenase activity in any of the BCG strains listed in this study (data not shown). The failure of BCG strains to utilize L- or D- alanine as the only nitrogen source for growth is due to the lack of a functional L-alanine dehydrogenase. To prove this, the *ald* gene [SEQ ID NO:1] coding for L-alanine dehydrogenase [SEQ ID NO:2] in the *M. tuberculosis* genome was cloned into a shuttle vector and transformed into BCG-Frappier and BCG-Pasteur. The resulting recombinant BCG strains were tested for their ability to grow in GAS medium containing L-alanine or D-alanine. Both recombinant strains, BCG-Frappier/*ald* and BCG-Pasteur/*ald*, grew rapidly in GAS medium containing either L-alanine or D-alanine (Fig. 6), while strains containing the cloning vector alone did not grow. This result shows that expression of a functional L-alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2] in BCG strains relieves the growth inhibition of BCG by L-alanine and D-alanine.

Example 5

Expressing L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6] in BCG relieves the inhibition of BCG growth by L-serine. L-Serine is used by *M. tuberculosis*, *M. avium* and *M. smegmatis*, but not *M. bovis* BCG, as the only nitrogen for growth. The failure of BCG to utilize L-serine as the only nitrogen source is likely to be caused by either mutations on or altered expression of the gene encoding L-serine dehydratase, *sdaA* [SEQ ID NO:5], in BCG. Expression of *sdaA* [SEQ ID NO:5; SEQ ID NO:6] of *M. tuberculosis* in BCG allows BCG strains to grow on L-serine as the only nitrogen source and relieves the inhibition of BCG growth by L-serine (Fig. 8).

Example 6

Inhibition of BCG growth by L-alanine, D-alanine and L-serine are likely to occur by blocking the activity of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO:14]. Glutamine synthetase plays a central role in nitrogen metabolism in bacteria (Reitzer, 1996). Working in tandem with glutamate synthase, glutamine synthetase catalyzes the synthesis of glutamine and glutamate, which together provide nitrogen for almost all amino acids, proteins, and nucleotides. In *Escherichia coli* and *Klebsiella aerogenes*,

glutamine synthetase is under feedback inhibition – purified glutamine synthetase is inhibited by L-alanine, L-serine and glycine (Reitzer, 1996). Glutamine synthetase was identified as an extracellular protein in *M. tuberculosis* and *M. bovis* BCG (Harth et al., 1994). It is likely that undegraded L-alanine inhibits glutamine synthetase and subsequently prevents the growth of BCG. If this were correct, then L-serine, which was not catabolized by BCG for growth (Table I), would also inhibit the growth of BCG by the same mechanism. Supporting this hypothesis, addition of L-serine to GAS medium containing only ammonium chloride as the nitrogen source inhibits the growth of BCG-Frappier, BCG-Pasteur or BCG-Japan (Fig. 7). Furthermore, if glutamine synthetase were the target of L-alanine and L-serine inhibition, then supplementing amino acids that can be converted to glutamate would also alleviate their effects, as demonstrated in *K. aerogenes* (Janes and Bender, 1998). Indeed, addition of L-glutamate and amino acids that could be catabolized to yield glutamate (L-glutamine, L-asparagine, and L-aspartate) allows the growth of BCG strains in the presence of alanine (Table I), but those that could not be catabolized to glutamate (e.g., L-lysine, L-methionine, L-leucine) fail to allow growth. BCG-Frappier and BCG-Pasteur are more sensitive than BCG-Japan to inhibition by alanine and serine, this is due to differences in the expression level or activity of glutamine synthetase [SEQ ID NO:7] to [SEQ ID NO:14], i.e., BCG-Japan produces more glutamine synthetase or with higher activity than BCG-Frappier or BCG-Pasteur.

The present invention has been described in detail and with particular reference to the preferred embodiments; however, it will be understood by one having ordinary skill in the art that changes can be made without departing from the spirit and scope thereof. For example, where the application refers to proteins, it is clear that peptides and polypeptides may often be used. Likewise, where a gene is described in the application, it is clear that nucleic acids or gene fragments may often be used.

All publications (including Genbank entries), patents and patent applications are incorporated by reference in their entirety to the same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated by reference in its entirety.

Table I

Comparative growth of *M. tuberculosis*, *M. smegmatis* and *M. bovis* BCG substrains in 7H9, Sauton, and glycerol-alanine-salts (GAS) medium.

Mycobacterium ^a	7H9	Sauton	GAS	GAS + L-Asn ^b	GAS + L-Asp ^b	GAS + L-Glu ^b	GAS + L-Gln ^b
<i>M. tuberculosis</i> ^c	+	+	+	+	+	+	+
<i>M. smegmatis</i>	+	+	+	+	+	+	+
BCG-Russia	+	+	+	+	+	+	+
BCG-Moreau	+	+	+	+	+	+	+
BCG-Japan	+	+	+	+	+	+	+
BCG-Sweden	+	+	+	+	+	+	+
BCG-Birkhaug	+	+	+	+	+	+	+
BCG-Prague	+	+	-	+	+	+	+
BCG-Glaxo	+	+	-	+	+	+	+
BCG-Denmark	+	+	-	+	+	+	+

RESULTS

BCG-Tice	+	+	-	+	+	+	+
BCG-Frappier	+	+	-	+	+	+	+
BCG-Phipps	+	+	-	+	+	+	+
BCG-Pasteur	+	+	-	+	+	+	+

^a Each 5 ml culture inoculated with 1×10⁷ cells of *M. smegmatis* or *M. bovis* BCG substrains.

^b L-Asn, L-Asp, L-Glu and L-Gln in GAS supplemented to a final concentration of 27 mM.

^c Based on research literature.

Table II

Comparative growth of *M. bovis* BCG-Japan, BCG-Frappier, BCG-Pasteur, *M. tuberculosis*, *M. avium* and *M. smegmatis*

Media ^a	BCG-Japan ^b	BCG-Frappier ^b	BCG-Pasteur ^b	<i>M. tuberculosis</i> ^c	<i>M. avium</i> ^c	<i>M. smegmatis</i> ^b
Sauton basal	-	-	-	-	-	-
Group 1						
Sauton + L-Asn	+++	+++	+++	+++	+++	+++
Sauton + L-Asp	+++	+++	+++	+++	+++	+++
Sauton + L-Glu	+++	+++	+++	+++	+++	+++
Sauton + L-Gln	+++	+++	+++	+++	+++	+++
Sauton + L-Cys	+++	+++	+++	+++	+++	+++
Sauton + NH ₄ Cl	+++	+++	+++	+++	+++	+++
Group 2						
Sauton + L-Arg	++	-	-	++	++	++
Sauton + L-His	++	-	-	++	++	++

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Sauton + L-Lys	++	-	NA	++	+++
Sauton + L-Pro	++	-	NA	-	+++
Sauton + GABA	++	-	NA	NA	+++
Sauton + L-Ornithine	++	-	NA	NA	+++
Group 3					
Sauton + L-Ala	-	-	+++	+++	+++
Sauton + L-Ser	-	-	+++	++	++
Sauton + L-Leu	-	-	+++	++	++
Sauton + L-Ile	-	-	+++	++	++
Sauton + L-Met	-	-	NA	++	++
Sauton + Glycine	-	-	+++	NA	+++
Group 4					
Sauton + L-Trp	-	-	-	-	-
Sauton + L-Phe	-	-	+++	-	-

RESULTS OBTAINED

Sauton + L-Tyr	-	-	-	-
Sauton + L-Val	-	-	NA	-
Sauton + L-Thr	-	-	NA	-

^a All amino acids, L-Ornithine and GABA supplemented to final concentration of 27mM. NH₄Cl was tested at 1mM, 27 mM and 96 mM.

^b Each 5 ml culture inoculated with 1x10⁷ cells of *M. smegmatis* or *M. bovis* BCG substrains.

^c Based on research literature.

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We claim:

1. A live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase activity [SEQ ID NO:7 to SEQ ID NO:14], or L-serine dehydratase activity [SEQ ID NO:5; SEQ ID NO:6].
2. A live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide selected from the group consisting of alanine dehydrogenase [SEQ ID NO:1; SEQ ID NO:2], glutamine synthetase [SEQ ID NO:7 to SEQ ID NO:14] and L-serine dehydratase [SEQ ID NO:5; SEQ ID NO:6].
3. A live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid comprises all or part of at least one nucleic acid molecule selected from the group consisting of [SEQ ID NO:1], [SEQ ID NO:2], [SEQ ID NO:5], [SEQ ID NO:6], [SEQ ID NO:7], [SEQ ID NO:8], [SEQ ID NO:9], [SEQ ID NO:10], [SEQ ID NO:11], [SEQ ID NO:12], [SEQ ID NO:13], and [SEQ ID NO:14].
4. The live recombinant *Mycobacterium bovis*-BCG strain of claim 1, 2 or 3 wherein the *Mycobacterium bovis*-BCG strain is selected from the group consisting of *Mycobacterium bovis*-BCG-Russia, *Mycobacterium bovis*-BCG-Moreau, *Mycobacterium bovis*-BCG-Japan, *Mycobacterium bovis*-BCG-Sweden, *Mycobacterium bovis*-BCG-Birkhaug, *Mycobacterium bovis*-BCG-Prague, *Mycobacterium bovis*-BCG-Glaxo, *Mycobacterium bovis*-BCG-Denmark, *Mycobacterium bovis*-BCG-Tice, *Mycobacterium bovis*-BCG-Frappier, *Mycobacterium bovis*-BCG-Connaught, *Mycobacterium bovis*-BCG-Phipps, and *Mycobacterium bovis*-BCG-Pasteur.
5. A pharmaceutical composition comprising the live recombinant *Mycobacterium bovis*-BCG strain of claim 1, 2 or 3.

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6. A vaccine or immunogenic composition for treatment or prophylaxis of a mammal against challenge by mycobacteria comprising the live recombinant *Mycobacterium bovis*-BCG strain of claim 1, 2 or 3.
7. The vaccine or immunogenic composition of claim 6 wherein the mycobacteria is *Mycobacterium tuberculosis*.
8. The vaccine or immunogenic composition of claim 6 or 7 further comprising a pharmaceutically acceptable carrier.
9. The vaccine or immunogenic composition of claim 6, 7, or 8 further comprising an adjuvant.
10. The vaccine or immunogenic composition of claim 6, 7, 8 or 9 further comprising immunogenic materials from one or more other pathogens.
11. A method for treatment or prophylaxis of a mammal against challenge by *Mycobacterium tuberculosis* or *Mycobacterium bovis* comprising administering to the mammal the vaccine or immunogenic composition of claim 1, 2 or 3.
12. The method of claim 11 wherein the mammal is a cow.
13. The method of claim 11 wherein the mammal is a human.
14. The method of claim 11 wherein the vaccine or immunogenic composition is administered in the presence of an adjuvant.
15. A method for treatment or prophylaxis of a mammal against cancer comprising administering to the mammal the vaccine or immunogenic composition of claim 1, 2 or 3.
16. The method of claim 15 wherein the vaccine or immunogenic composition is administered in the presence of an adjuvant.
17. The method of claim 15 or 16 wherein the cancer is bladder cancer.

18. A test kit comprising the live recombinant *Mycobacterium bovis*-BCG strain of claim 1, 2 or 3.
19. A media composition for inhibiting the growth of *Mycobacterium bovis*-BCG comprising alanine as the only nitrogen source for growth.
20. A media composition for inhibiting the growth of *Mycobacterium bovis*-BCG comprising serine as the only nitrogen source for growth.
21. The media composition of claim 19 or 20 further comprising:
 - (a) a carbon source;
 - (b) iron;
 - (c) magnesium; and
 - (d) SO₄.
22. A media composition of claim 21 wherein the carbon source is selected from the group consisting of glycerol, dextrose, citrate and glucose.
23. A method for inhibiting the growth of *Mycobacterium bovis*-BCG comprising:
 - (a) obtaining a sample comprising *Mycobacterium*; and
 - (b) culturing the sample in a selective media.
24. The method of claim 23, wherein the selective media comprises alanine as the only nitrogen source for growth.
25. The method of claim 23, wherein the selective media comprises serine as the only nitrogen source for growth.
26. A method of culturing *Mycobacterium bovis*-BCG comprising:
 - (a) obtaining a sample of *Mycobacterium*; and

(b) culturing the sample in differential media.

27. The method of claim 26, wherein the differential media comprises histidine.

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Abstract

The invention relates to a live recombinant *Mycobacterium bovis*-BCG strain comprising a nucleic acid capable of expression, the nucleic acid encoding at least one protein or polypeptide that exhibits alanine dehydrogenase activity, glutamine synthetase activity, or serine dehydratase activity.

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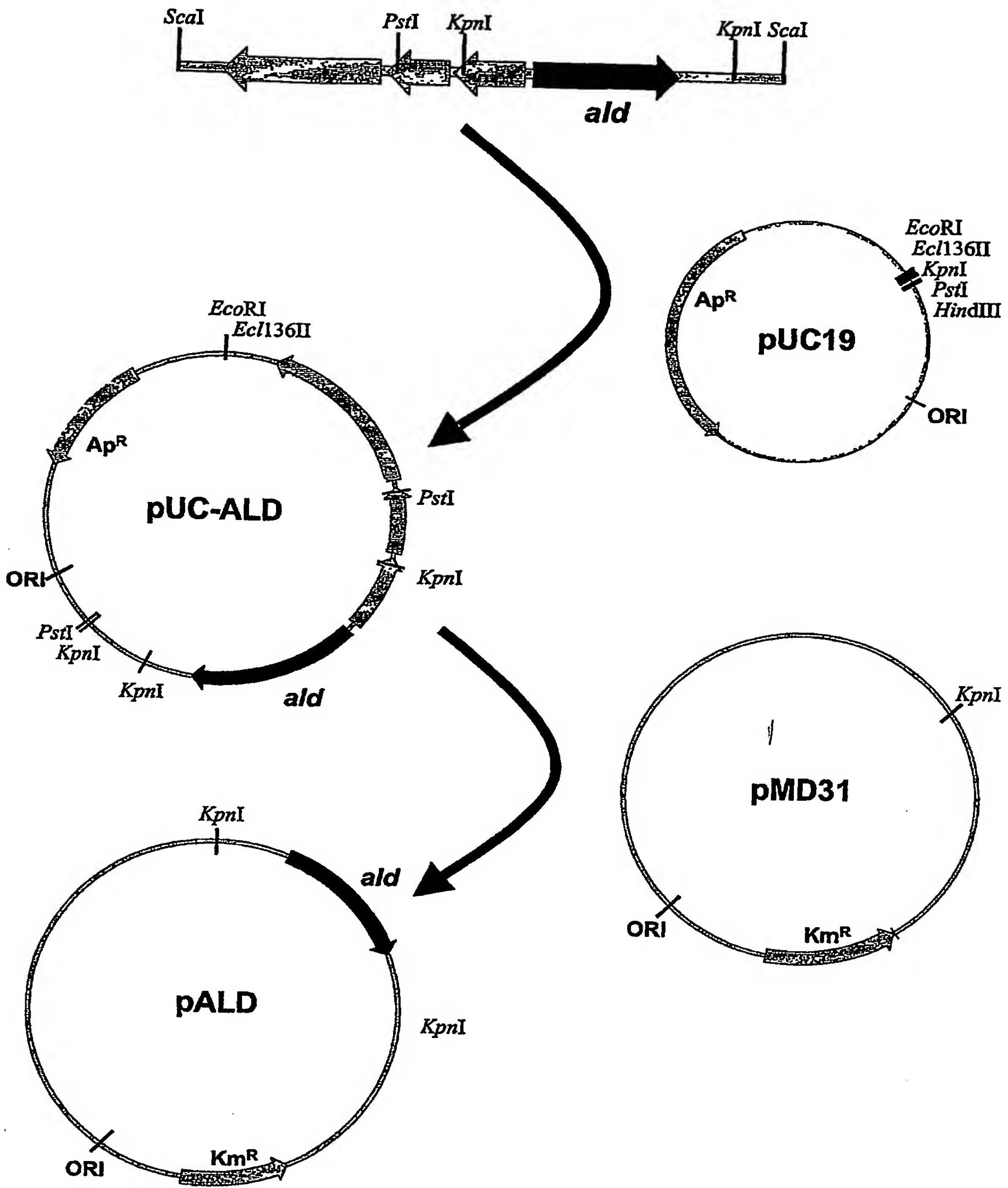


Fig. 1

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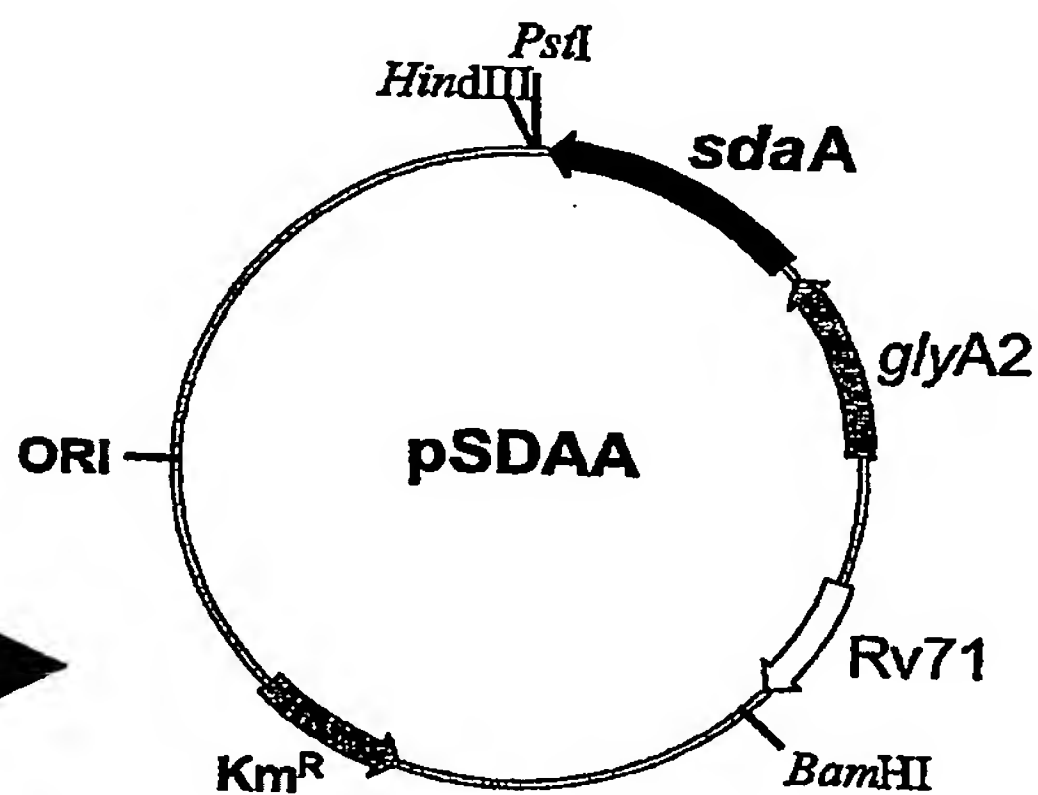
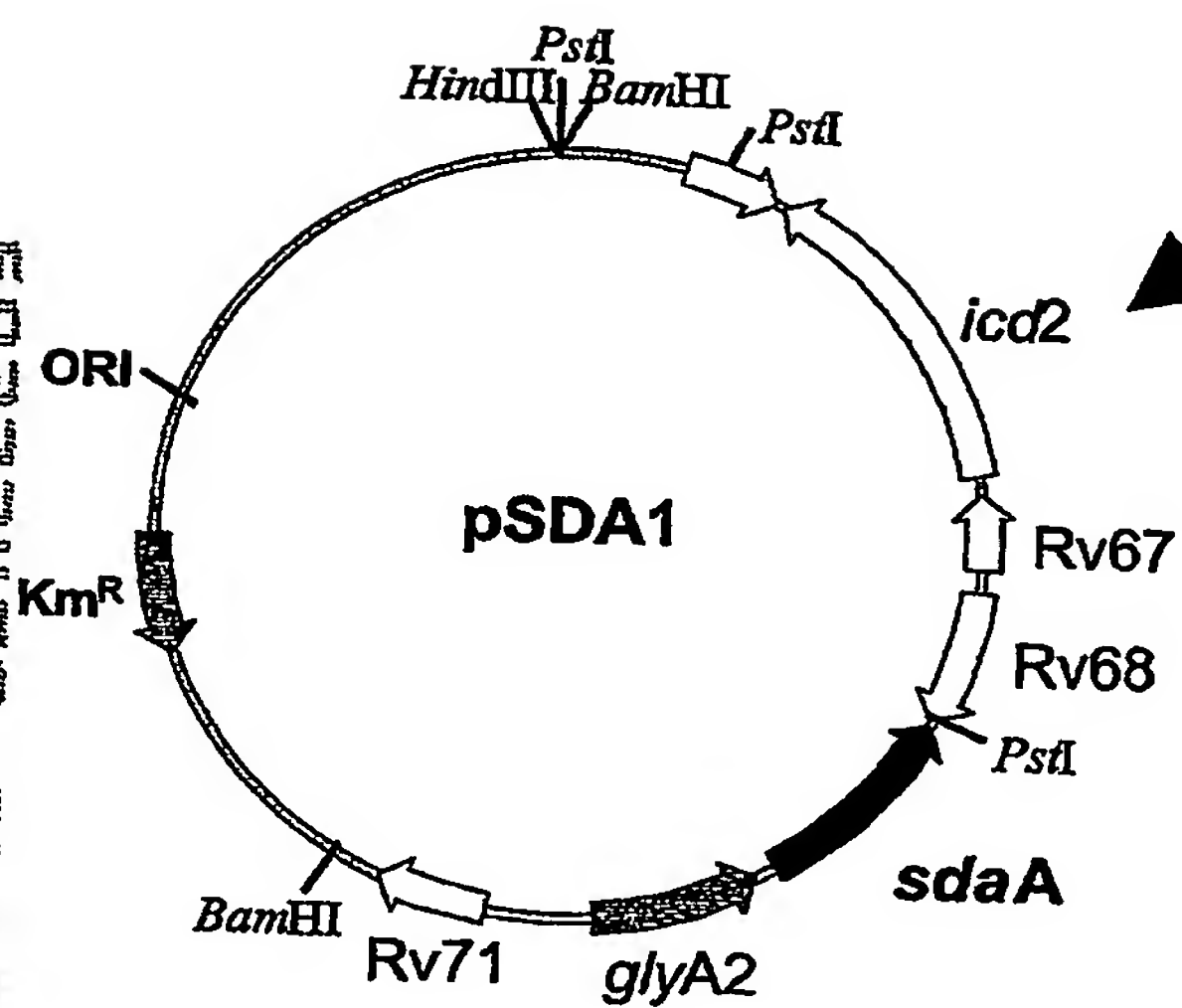
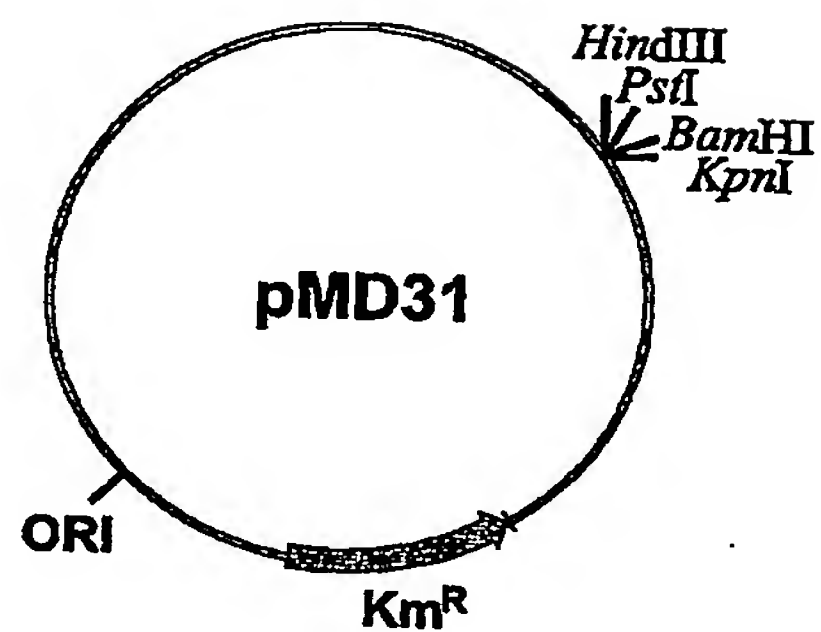
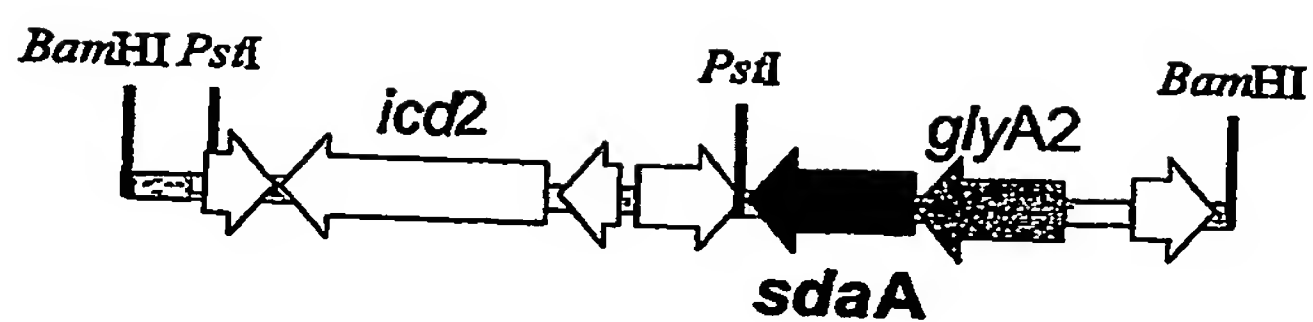


Fig. 2

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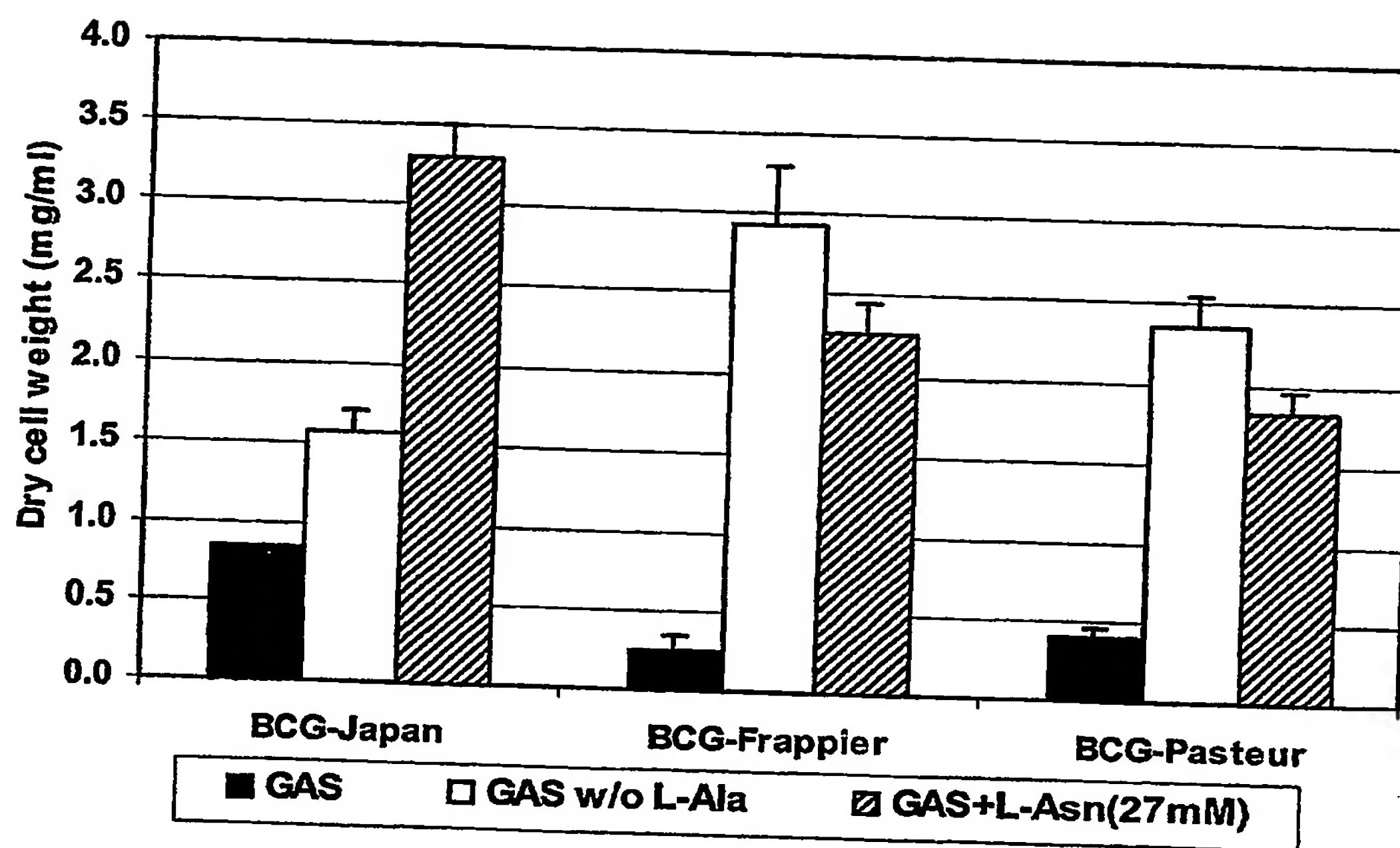
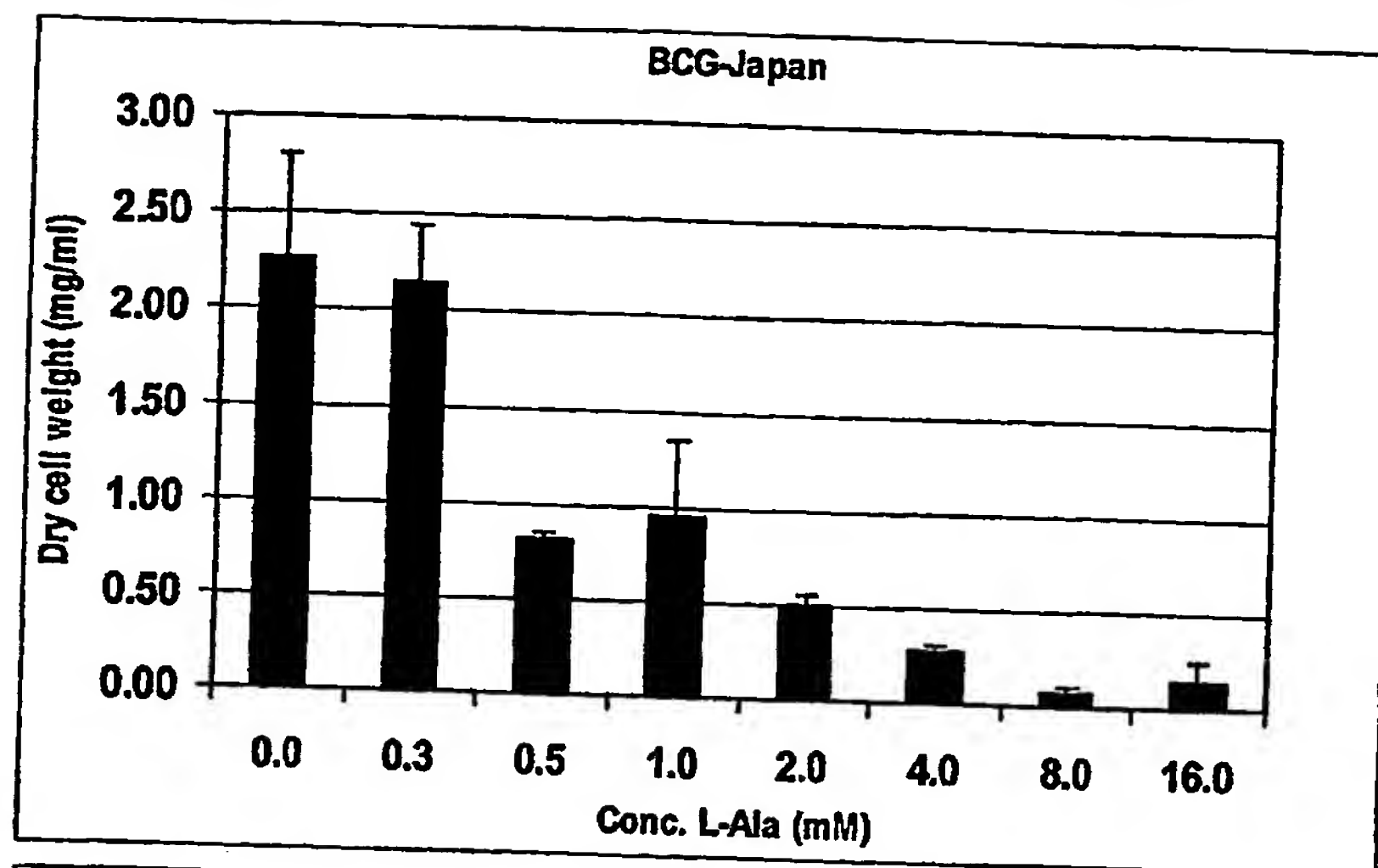
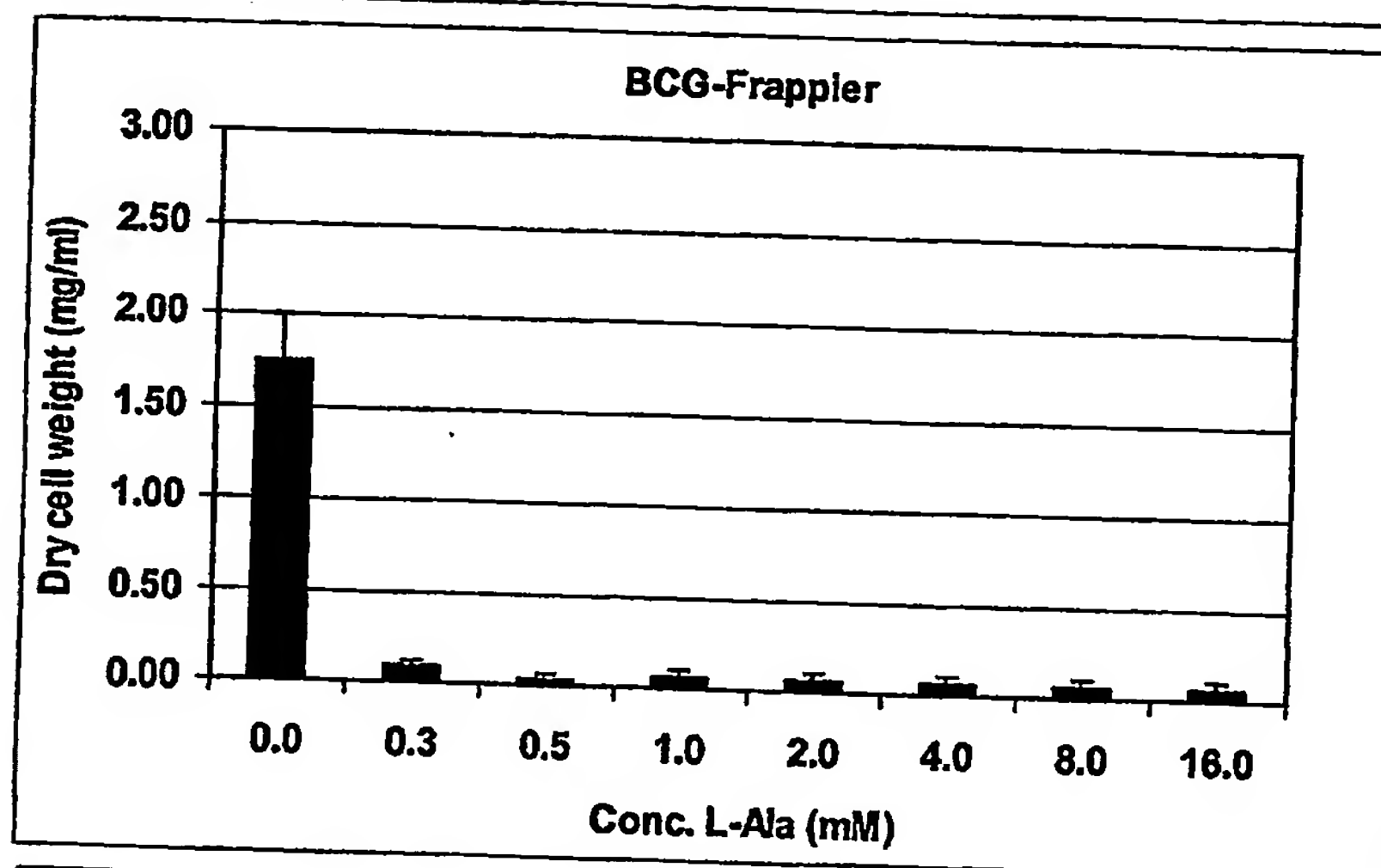


Fig. 3

a)



b)



c)

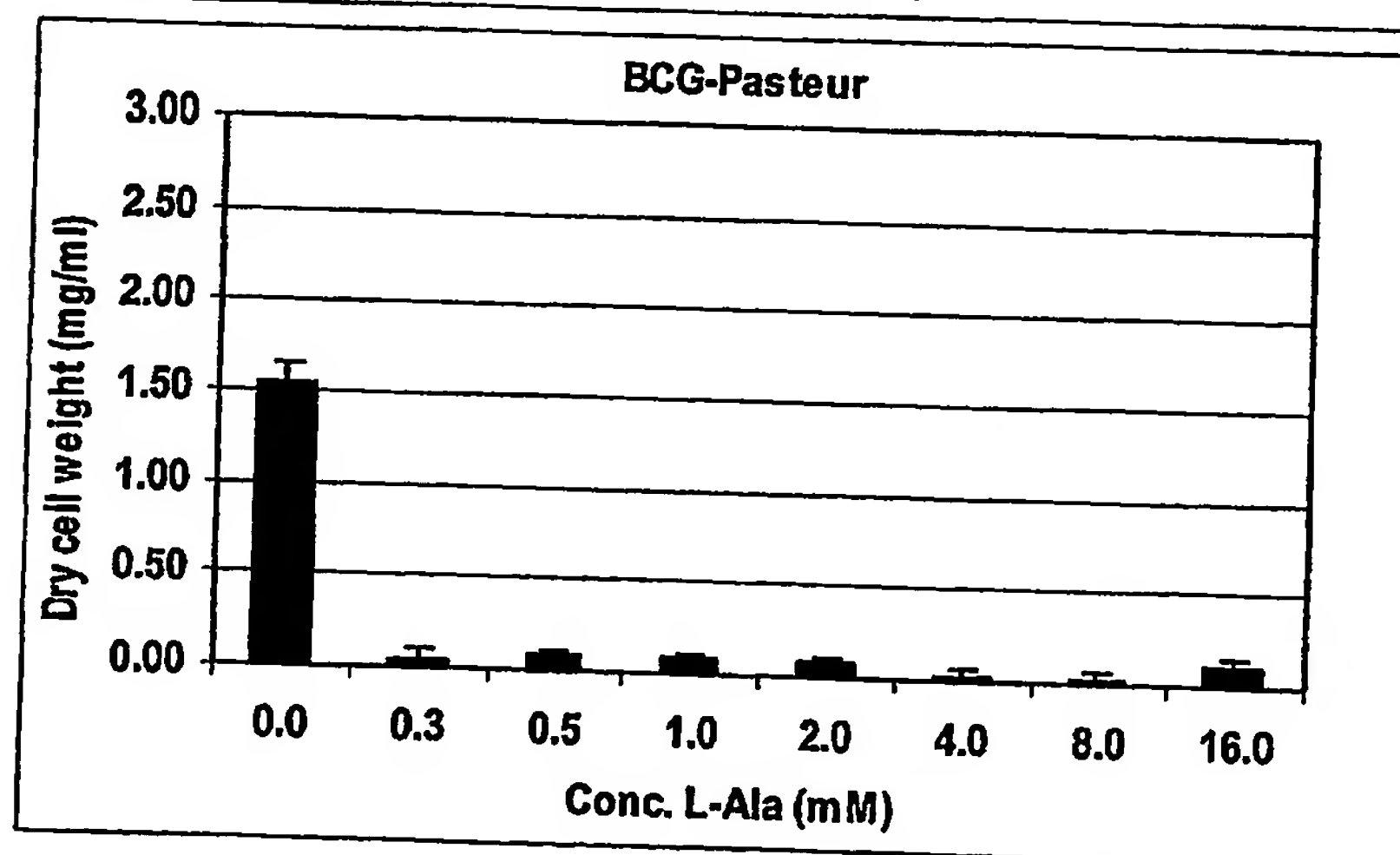


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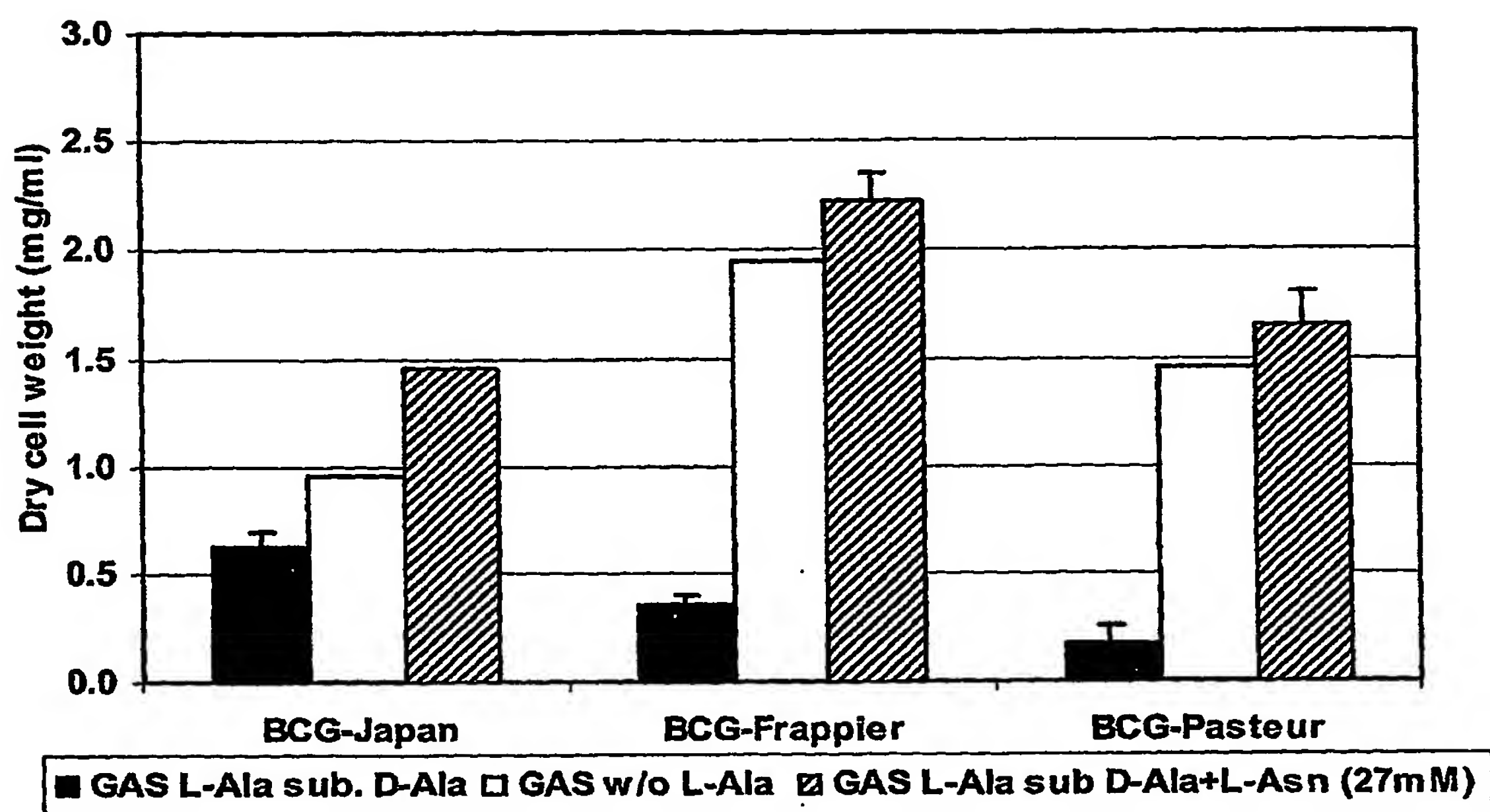


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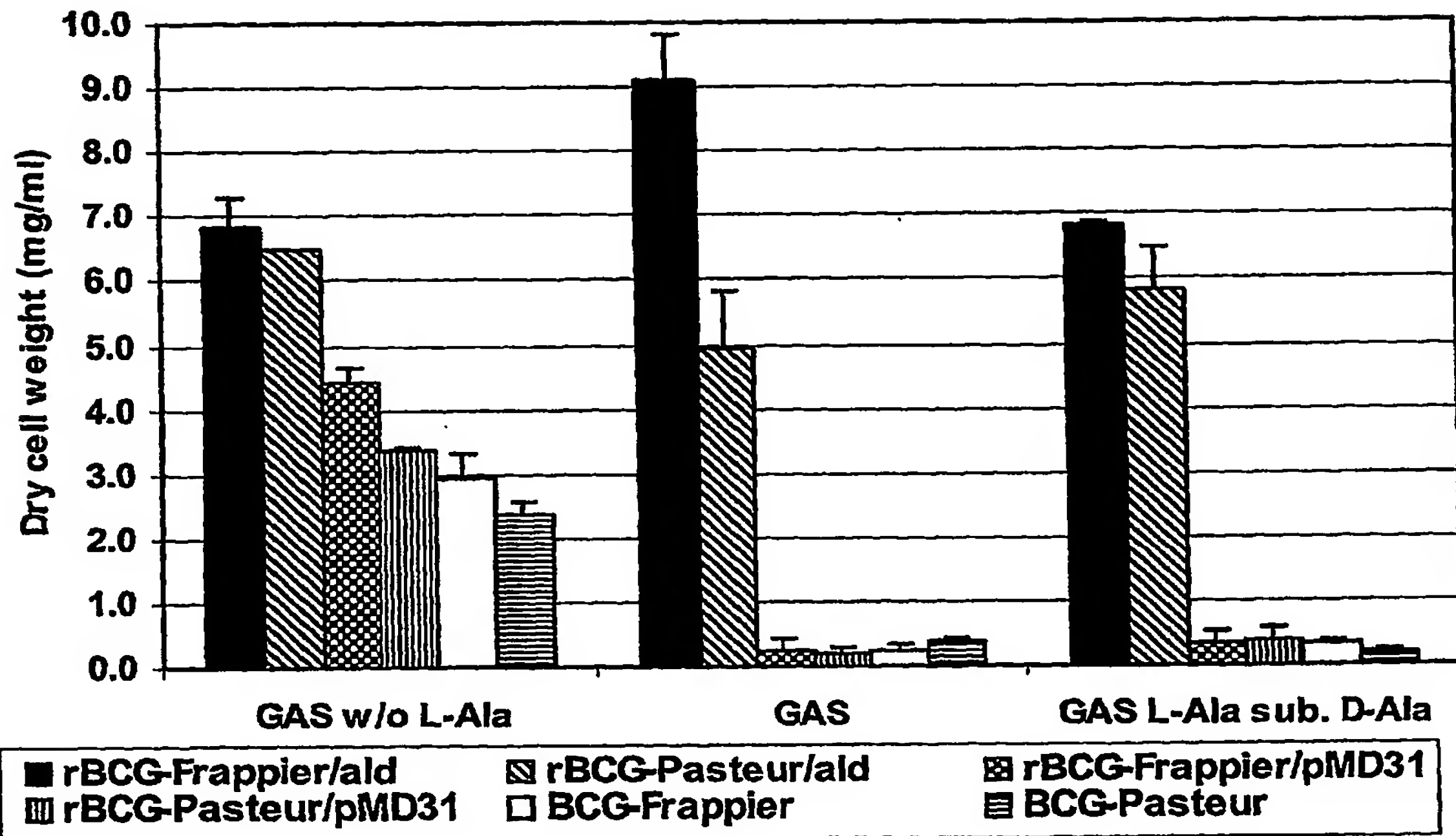


Fig. 6

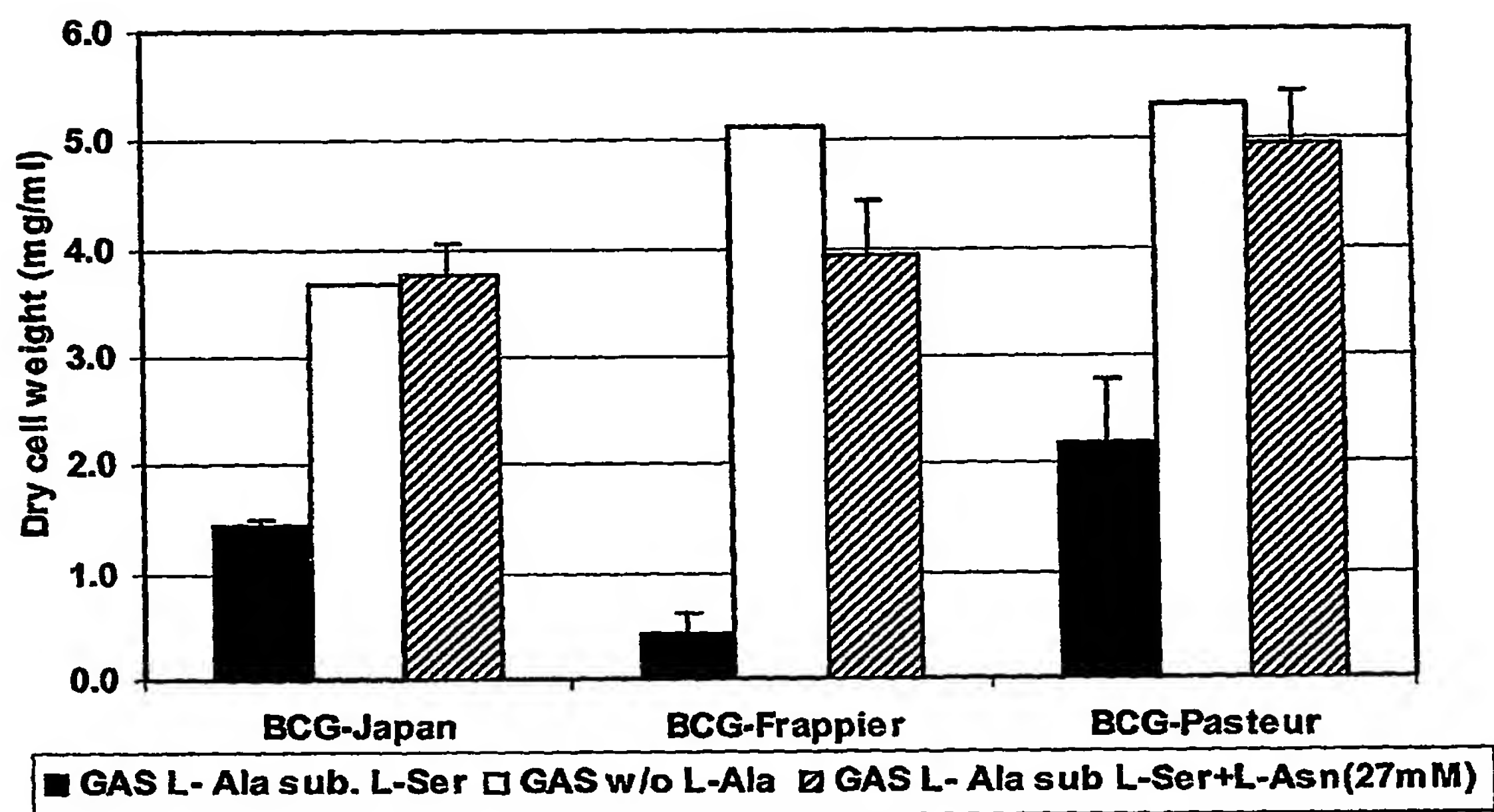


Fig. 7

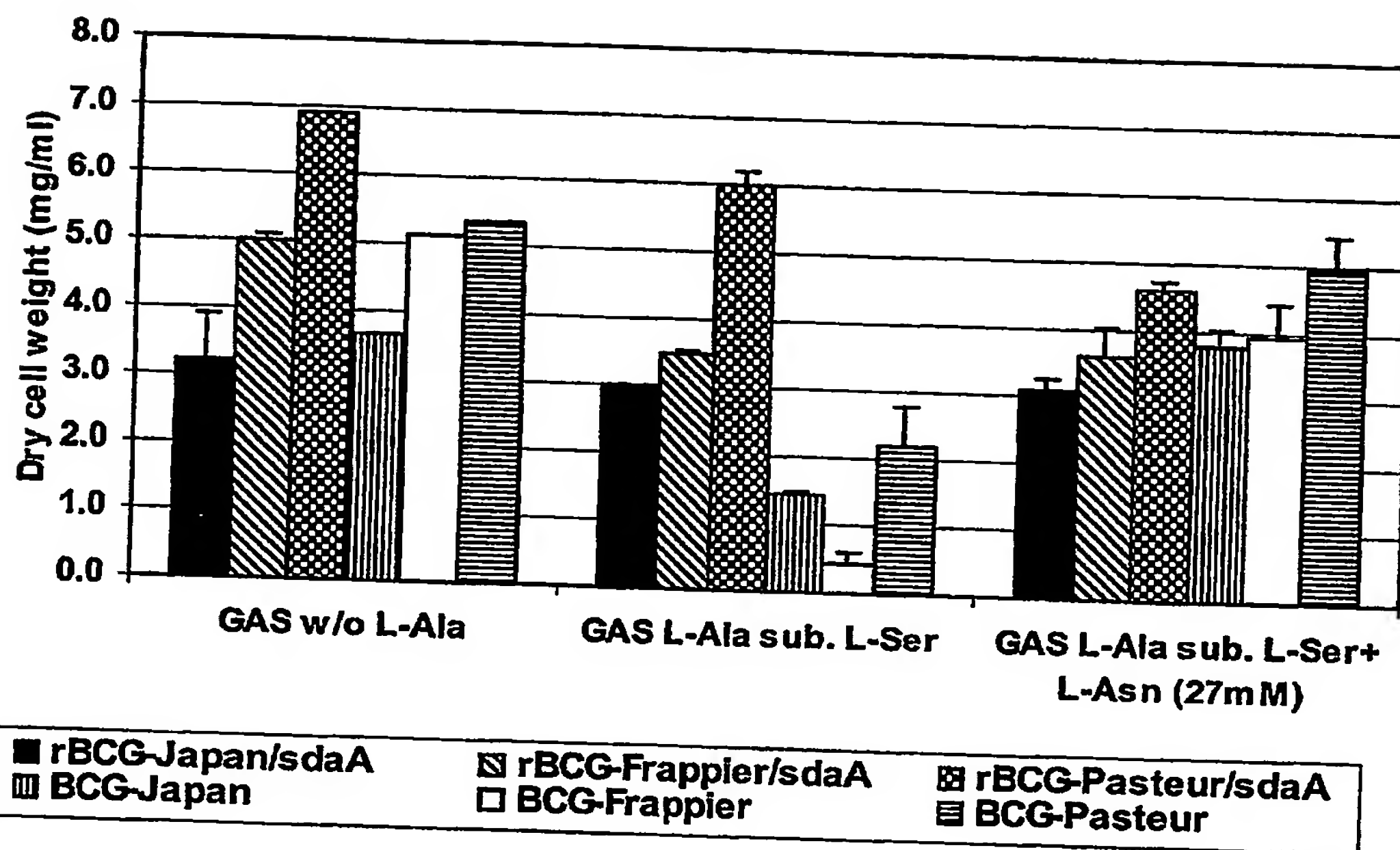


Fig. 8

<i>M. tb</i>	ATG	CGC	GTC	GGT	ATT	COG	ACC	GAG	ACC	AAA	AAC	AAC	GAA	TTC	CGG	GTG	GCC	ATC
<i>M. bovis</i>	ATG	CGC	GTC	GGT	ATT	COG	ACC	GAG	ACC	AAA	AAC	AAC	GAA	TTC	CGG	GTG	GCC	ATC
<i>M. tb</i>	ACC	CCG	GCC	GGC	GTC	GCG	GAA	CTA	ACC	CGT	CGT	GGC	CAT	GAG	GTG	CTC	ATC	CAG
<i>M. bovis</i>	ACC	CCG	GCC	GGC	GTC	GCG	GAA	CTA	ACC	CGT	CGT	GGC	CAT	GAG	GTG	CTC	ATC	CAG
<i>M. tb</i>	GCA	GGT	GCC	GGA	GAG	GGC	TCG	GCT	ATC	ACC	GAC	GCG	GAT	TTC	AAG	GCG	GCA	GGC
<i>M. bovis</i>	GCA	GGT	GCC	GGA	GAG	GGC	TCG	GCT	ATC	ACC	GAC	GCG	GAT	TTC	AAG	GCG	GCA	GGC
<i>M. tb</i>	GCG	CAA	CTG	GTC	GGC	ACC	GCC	GAC	CAG	GTG	TGG	GCC	GAC	GCT	GAT	TTA	TTG	CTC
<i>M. bovis</i>	GCG	CAA	CTG	GTC	GGC	ACC	GCC	GAC	CAG	GTG	TGG	GCC	GAC	GCT	GAT	TTA	TTG	CTC
<i>M. tb</i>	AAG	GTC	AAA	GAA	CCG	ATA	GCG	GCG	GAA	TAC	GGC	CGC	CTG	CGA	CAC	GGG	CAG	ATC
<i>M. bovis</i>	AAG	GTC	AAA	GAA	CCG	ATA	GCG	GCG	GAA	TAC	GGC	CGC	CTG	CGA	CAC	GGG	CAG	ATC

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M.bovis TGT TCA CGT TCT TGC ATT TGG CCG CGT CAC GTG CTT GCA CCG ATG CGT TGT TGG

***M.tb* GAT TCC GGC ACC ACG TCA ATT GCC TAC GAG ACC GTC CAG ACC GCC GAC GGC GCA**
***M.bovis* ATT CCG GCA CCA CGT CAA TTG CCT ACG AGA CCG TCC AGA CCG CCG ACG GCG CAC**

M. tb CTA CCC CTG CTT GCC CCG ATG AGC GAA GTC GCC GGT CGA CTC GCC GCC CAG GTT
M. bovis TAC CCC TGC TTG CCC CGA TGA

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M.tb RHTRYSSAYELEGAVKRADLVIGAVLVPGAKAPKLVSNLVAHMKPGAVLVDIAIDQGGCFEGSRPTTYD

M. lb HPTFAVHDTLFYCVANMPASVPKSTYALTNATMPYVLELADHGWRAACRSNPALAKGLSTHEGALLSERV

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Fig. 9

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Ala Ile Thr Pro Ala Gly Val Ala Glu Leu Thr Arg Arg Gly His Glu
 20 25 30

Val Leu Ile Gln Ala Gly Ala Gly Glu Gly Ser Ala Ile Thr Asp Ala
 35 40 45

Asp Phe Lys Ala Ala Gly Ala Gln Leu Val Gly Thr Ala Asp Gln Val
 50 55 60

Trp Ala Asp Ala Asp Leu Leu Leu Lys Val Lys Glu Pro Ile Ala Ala
 65 70 75 80

Glu Tyr Gly Arg Leu Arg His Gly Gln Ile Leu Phe Thr Phe Leu His
 85 90 95

Leu Ala Ala Ser Arg Ala Cys Thr Asp Ala Leu Leu Asp Ser Gly Thr
 100 105 110

Thr Ser Ile Ala Tyr Glu Thr Val Gln Thr Ala Asp Gly Ala Leu Pro
 115 120 125

Leu Leu Ala Pro Met Ser Glu Val Ala Gly Arg Leu Ala Ala Gln Val
 130 135 140

Gly Ala Tyr His Leu Met Arg Thr Gln Gly Gly Arg Gly Val Leu Met
 145 150 155 160

209140-0542E09

Gly Gly Val Pro Gly Val Glu Pro Ala Asp Val Val Val Ile Gly Ala
165 170 175

Gly Thr Ala Gly Tyr Asn Ala Ala Arg Ile Ala Asn Gly Met Gly Ala
180 185 190

Thr Val Thr Val Leu Asp Ile Asn Ile Asp Lys Leu Arg Gln Leu Asp
195 200 205

Ala Glu Phe Cys Gly Arg Ile His Thr Arg Tyr Ser Ser Ala Tyr Glu
210 215 220

Leu Glu Gly Ala Val Lys Arg Ala Asp Leu Val Ile Gly Ala Val Leu
225 230 235 240

Val Pro Gly Ala Lys Ala Pro Lys Leu Val Ser Asn Ser Leu Val Ala
245 250 255

His Met Lys Pro Gly Ala Val Leu Val Asp Ile Ala Ile Asp Gln Gly
260 265 270

Gly Cys Phe Glu Gly Ser Arg Pro Thr Thr Tyr Asp His Pro Thr Phe
275 280 285

Ala Val His Asp Thr Leu Phe Tyr Cys Val Ala Asn Met Pro Ala Ser
290 295 300

Val Pro Lys Thr Ser Thr Tyr Ala Leu Thr Asn Ala Thr Met Pro Tyr
305 310 315 320

Val Leu Glu Leu Ala Asp His Gly Trp Arg Ala Ala Cys Arg Ser Asn
325 330 335

Pro Ala Leu Ala Lys Gly Leu Ser Thr His Glu Gly Ala Leu Leu Ser
340 345 350

Glu Arg Val Ala Thr Asp Leu Gly Val Pro Phe Thr Glu Pro Ala Ser
355 360 365

Val Leu Ala
370

<210> 3
 <211> 399
 <212> DNA
 <213> Mycobacterium bovis

<220>
 <221> CDS
 <222> (1)..(399)

<400> 3
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 Met Arg Val Gly Ile Pro Thr Glu Thr Lys Asn Asn Glu Phe Arg Val
 1 5 10 15
 gcc atc acc ccg gcc ggc gtc gcg gaa cta acc cgt cgt ggc cat gag 96
 Ala Ile Thr Pro Ala Gly Val Ala Glu Leu Thr Arg Arg Gly His Glu
 20 25 30
 gtg ctc atc cag gca ggt gcc gga gag ggc tcg gct atc acc gac gcg 144
 Val Leu Ile Gln Ala Gly Ala Gly Glu Gly Ser Ala Ile Thr Asp Ala
 35 40 45
 gat ttc aag gcg gca ggc gcg caa ctg gtc ggc acc gcc gac cag gtg 192
 Asp Phe Lys Ala Ala Gly Ala Gln Leu Val Gly Thr Ala Asp Gln Val
 50 55 60
 tgg gcc gac gct gat tta ttg ctc aag gtc aaa gaa ccg ata gcg gcg 240
 Trp Ala Asp Ala Asp Leu Leu Lys Val Lys Glu Pro Ile Ala Ala
 65 70 75 80
 gaa tac ggc cgc ctg cga cac ggg cga tct tgt tca cgt tct tgc att 288
 Glu Tyr Gly Arg Leu Arg His Gly Arg Ser Cys Ser Arg Ser Cys Ile
 85 90 95
 tgg ccg cgt cac gtg ctt gca ccg atg cgt tgt tgg att ccg gca cca 336
 Trp Pro Arg His Val Leu Ala Pro Met Arg Cys Trp Ile Pro Ala Pro
 100 105 110
 cgt caa ttg cct acg aga ccg tcc aga ccg ccg acg gcg cac tac ccc 384
 Arg Gln Leu Pro Thr Arg Pro Ser Arg Pro Pro Thr Ala His Tyr Pro
 115 120 125
 tgc ttg ccc cga tga 399
 Cys Leu Pro Arg
 130

<210> 4
 <211> 132
 <212> PRT
 <213> Mycobacterium bovis

<400> 4
 Met Arg Val Gly Ile Pro Thr Glu Thr Lys Asn Asn Glu Phe Arg Val
 1 5 10 15

Ala Ile Thr Pro Ala Gly Val Ala Glu Leu Thr Arg Arg Gly His Glu
20 25 30

Val Leu Ile Gln Ala Gly Ala Gly Glu Gly Ser Ala Ile Thr Asp Ala
35 40 45

Asp Phe Lys Ala Ala Gly Ala Gln Leu Val Gly Thr Ala Asp Gln Val
50 55 60

Trp Ala Asp Ala Asp Leu Leu Leu Lys Val Lys Glu Pro Ile Ala Ala
65 70 75 80

Glu Tyr Gly Arg Leu Arg His Gly Arg Ser Cys Ser Arg Ser Cys Ile
85 90 95

Trp Pro Arg His Val Leu Ala Pro Met Arg Cys Trp Ile Pro Ala Pro
100 105 110

Arg Gln Leu Pro Thr Arg Pro Ser Arg Pro Pro Thr Ala His Tyr Pro
115 120 125

Cys Leu Pro Arg
130

<210> 5
<211> 1386
<212> DNA
<213> Mycobacterium tuberculosis

<220>
<221> CDS
<222> (1)..(1386)
<223> Sequence is identical to the complement of nucleotides 13172-14551
of GenBank entry GB:MTV030 [AL021428]
Sequence is identical to the complement of nucleotides 13195-14580
of GenBank entry GB:AE006919

<400> 5
atg acc atc agc gtc ttc gac ctg ttc acc atc ggc atc ggg ccg tcc 48
Met Thr Ile Ser Val Phe Asp Leu Phe Thr Ile Gly Ile Gly Pro Ser
1 5 10 15

agt tcc cac acc gtg gga ccg atg cgc gcg gca aac cag ttc gta gtt 96
Ser Ser His Thr Val Gly Pro Met Arg Ala Ala Asn Gln Phe Val Val
20 25 30

gcg ctg cgc cgc cgg ggc cac ctg gat gac ctc gag gcg atg cga gtg 144

Ala	Leu	Arg	Arg	Arg	Gly	His	Leu	Asp	Asp	Leu	Glu	Ala	Met	Arg	Val	
	35						40					45				
gat	ctg	ttc	ggc	tcg	ctc	gcg	gcc	acc	gga	gcc	ggc	cat	ggc	acc	atg	192
Asp	Leu	Phe	Gly	Ser	Leu	Ala	Ala	Thr	Gly	Ala	Gly	His	Gly	Thr	Met	
	50					55					60					
tcg	gcg	ata	ttg	ctg	ggg	ctg	gaa	ggc	tgc	cag	cca	gaa	acg	att	acc	240
Ser	Ala	Ile	Leu	Leu	Gly	Leu	Glu	Gly	Cys	Gln	Pro	Glu	Thr	Ile	Thr	
65					70				75						80	
acc	gaa	cac	aag	gaa	cgc	cgg	ctc	gcc	gag	atc	gca	gcg	tcc	ggc	gtg	288
Thr	Glu	His	Lys	Glu	Arg	Arg	Leu	Ala	Glu	Ile	Ala	Ala	Ser	Gly	Val	
				85					90					95		
acg	cga	atc	ggc	ggc	gtc	att	ccg	gtc	ccg	ctg	acc	gag	cgt	gat	atc	336
Thr	Arg	Ile	Gly	Gly	Val	Ile	Pro	Val	Pro	Leu	Thr	Glu	Arg	Asp	Ile	
			100					105						110		
gac	ctg	cat	ccc	gac	atc	gtt	ctg	cca	acg	cat	ccc	aac	gga	atg	acg	384
Asp	Leu	His	Pro	Asp	Ile	Val	Leu	Pro	Thr	His	Pro	Asn	Gly	Met	Thr	
		115					120					125				
ttc	act	gcc	gcg	ggc	cca	cac	ggc	cgc	gtc	ttg	gcc	acc	gag	act	tat	432
Phe	Thr	Ala	Ala	Gly	Pro	His	Gly	Arg	Val	Leu	Ala	Thr	Glu	Thr	Tyr	
	130					135					140					
ttt	tcg	gtg	ggc	gga	ggg	ttc	atc	gtc	acg	gaa	cag	acc	agc	ggc	aac	480
Phe	Ser	Val	Gly	Gly	Gly	Phe	Ile	Val	Thr	Glu	Gln	Thr	Ser	Gly	Asn	
145					150					155					160	
agc	ggc	caa	cat	cca	tgc	tca	gtt	gcc	ctt	ccc	tac	gtg	tcg	gcc	caa	528
Ser	Gly	Gln	His	Pro	Cys	Ser	Val	Ala	Leu	Pro	Tyr	Val	Ser	Ala	Gln	
				165				170						175		
gaa	ctg	ctg	gac	atc	tgt	gac	cgc	ctc	gac	gtg	tca	att	agc	gaa	gcg	576
Glu	Leu	Leu	Asp	Ile	Cys	Asp	Arg	Leu	Asp	Val	Ser	Ile	Ser	Glu	Ala	
			180					185					190			
gcg	ctg	cgc	aac	gaa	aca	tgt	tgc	cgc	acc	gag	aac	gag	gta	cgc	gcc	624
Ala	Leu	Arg	Asn	Glu	Thr	Cys	Cys	Arg	Thr	Glu	Asn	Glu	Val	Arg	Ala	
		195				200						205				
gcg	ctg	ctg	cac	ctg	cgc	gac	gtc	atg	gtt	gag	tgc	gaa	cag	cgg	agc	672
Ala	Leu	Leu	His	Leu	Arg	Asp	Val	Met	Val	Glu	Cys	Glu	Gln	Arg	Ser	
	210					215					220					
atc	gct	cgc	gaa	ggg	ttg	ctt	cct	ggc	ggc	ctc	cgg	gtg	cgc	cgg	cga	720
Ile	Ala	Arg	Glu	Gly	Leu	Leu	Pro	Gly	Gly	Leu	Arg	Val	Arg	Arg	Arg	
225					230					235					240	
gcg	aag	gtg	tgg	tat	gac	cgc	ttg	aac	gcc	gaa	gac	ccc	act	cgc	aag	768
Ala	Lys	Val	Trp	Tyr	Asp	Arg	Leu	Asn	Ala	Glu	Asp	Pro	Thr	Arg	Lys	
			245					250						255		
ccg	gaa	ttc	gct	gag	gac	tgg	gtc	aac	ctg	gtc	gcg	ctg	gca	gtc	aac	816
Pro	Glu	Phe	Ala	Glu	Asp	Trp	Val	Asn	Leu	Val	Ala	Leu	Ala	Val	Asn	

260	265	270	
gag gag aac gcc tcc ggt ggg cgc gtc gtc acc gcc ccg acc aac ggt Glu Glu Asn Ala Ser Gly Gly Arg Val Val Thr Ala Pro Thr Asn Gly 275 280 285			864
gcc gcc ggc atc gtg ccg gcg gtc ctg cac tac gca atc cac tac acg Ala Ala Gly Ile Val Pro Ala Val Leu His Tyr Ala Ile His Tyr Thr 290 295 300			912
tcg gcc ggc gcg ggg gac ccc gac gat gtc acc gtg cga ttc ctg ctc Ser Ala Gly Ala Gly Asp Pro Asp Asp Val Thr Val Arg Phe Leu Leu 305 310 315 320			960
act gct gga gcc atc gga tcg ttg ttc aag gag cga gca tcg atc tcc Thr Ala Gly Ala Ile Gly Ser Leu Phe Lys Glu Arg Ala Ser Ile Ser 325 330 335			1008
gga gcc gag gtc ggc tgt cag ggc gag gtc ggc tcc gcg gcc gcc atg Gly Ala Glu Val Gly Cys Gln Gly Glu Val Gly Ser Ala Ala Ala Met 340 345 350			1056
gcc gcc gcc gga ttg gct gaa atc ctc ggc ggc aca ccg cga caa gtg Ala Ala Ala Gly Leu Ala Glu Ile Leu Gly Gly Thr Pro Arg Gln Val 355 360 365			1104
gaa aac gcc gcc gag atc gcc atg gaa cac agc ctc ggc ctg acc tgt Glu Asn Ala Ala Glu Ile Ala Met Glu His Ser Leu Gly Leu Thr Cys 370 375 380			1152
gac ccc atc gcc ggg ctg gtg cag atc ccc tgc atc gaa cgc aac gcg Asp Pro Ile Ala Gly Leu Val Gln Ile Pro Cys Ile Glu Arg Asn Ala 385 390 395 400			1200
att tcc gcc ggc aag gcc atc aac gcc gca cgg atg gca ttg cgc ggc Ile Ser Ala Gly Lys Ala Ile Asn Ala Ala Arg Met Ala Leu Arg Gly 405 410 415			1248
gac ggc atc cat cgc gtc acc ctc gac cag gtc atc gac acc atg cgc Asp Gly Ile His Arg Val Thr Leu Asp Gln Val Ile Asp Thr Met Arg 420 425 430			1296
gcc acc ggc gcg gac atg cac acc aag tac aag gaa acc tcg gcc ggc Ala Thr Gly Ala Asp Met His Thr Lys Tyr Lys Glu Thr Ser Ala Gly 435 440 445			1344
ggg ctc gcc atc aac gtc gca gtc aac atc gtc gag tgt tga Gly Leu Ala Ile Asn Val Ala Val Asn Ile Val Glu Cys 450 455 460			1386

<210> 6
 <211> 461
 <212> PRT
 <213> Mycobacterium tuberculosis
 <220>

<221>

<222>

<223> Sequence is identical to SwissProt entry SP:SDHL_MYCTU

Sequence is identical to GenBank entries GP:AE006919_13 and GP:MTV030_11

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Ser Ser His Thr Val Gly Pro Met Arg Ala Ala Asn Gln Phe Val Val
20 25 30

Ala Leu Arg Arg Arg Gly His Leu Asp Asp Leu Glu Ala Met Arg Val
35 40 45

Asp Leu Phe Gly Ser Leu Ala Ala Thr Gly Ala Gly His Gly Thr Met
50 55 60

Ser Ala Ile Leu Leu Gly Leu Glu Gly Cys Gln Pro Glu Thr Ile Thr
65 70 75 80

Thr Glu His Lys Glu Arg Arg Leu Ala Glu Ile Ala Ala Ser Gly Val
85 90 95

Thr Arg Ile Gly Gly Val Ile Pro Val Pro Leu Thr Glu Arg Asp Ile
100 105 110

Asp Leu His Pro Asp Ile Val Leu Pro Thr His Pro Asn Gly Met Thr
115 120 125

Phe Thr Ala Ala Gly Pro His Gly Arg Val Leu Ala Thr Glu Thr Tyr
130 135 140

Phe Ser Val Gly Gly Gly Phe Ile Val Thr Glu Gln Thr Ser Gly Asn
145 150 155 160

Ser Gly Gln His Pro Cys Ser Val Ala Leu Pro Tyr Val Ser Ala Gln
165 170 175

Glu Leu Leu Asp Ile Cys Asp Arg Leu Asp Val Ser Ile Ser Glu Ala
180 185 190

Ala Leu Arg Asn Glu Thr Cys Cys Arg Thr Glu Asn Glu Val Arg Ala
195 200 205

Ala Leu Leu His Leu Arg Asp Val Met Val Glu Cys Glu Gln Arg Ser
 210 215 220

Ile Ala Arg Glu Gly Leu Leu Pro Gly Gly Leu Arg Val Arg Arg Arg
 225 230 235 240

Ala Lys Val Trp Tyr Asp Arg Leu Asn Ala Glu Asp Pro Thr Arg Lys
 245 250 255

Pro Glu Phe Ala Glu Asp Trp Val Asn Leu Val Ala Leu Ala Val Asn
 260 265 270

Glu Glu Asn Ala Ser Gly Gly Arg Val Val Thr Ala Pro Thr Asn Gly
 275 280 285

Ala Ala Gly Ile Val Pro Ala Val Leu His Tyr Ala Ile His Tyr Thr
 290 295 300

Ser Ala Gly Ala Gly Asp Pro Asp Asp Val Thr Val Arg Phe Leu Leu
 305 310 315 320

Thr Ala Gly Ala Ile Gly Ser Leu Phe Lys Glu Arg Ala Ser Ile Ser
 325 330 335

Gly Ala Glu Val Gly Cys Gln Gly Glu Val Gly Ser Ala Ala Ala Met
 340 345 350

Ala Ala Ala Gly Leu Ala Glu Ile Leu Gly Gly Thr Pro Arg Gln Val
 355 360 365

Glu Asn Ala Ala Glu Ile Ala Met Glu His Ser Leu Gly Leu Thr Cys
 370 375 380

Asp Pro Ile Ala Gly Leu Val Gln Ile Pro Cys Ile Glu Arg Asn Ala
 385 390 395 400

Ile Ser Ala Gly Lys Ala Ile Asn Ala Ala Arg Met Ala Leu Arg Gly
 405 410 415

Asp Gly Ile His Arg Val Thr Leu Asp Gln Val Ile Asp Thr Met Arg
 420 425 430

Ala Thr Gly Ala Asp Met His Thr Lys Tyr Lys Glu Thr Ser Ala Gly
435 440 445

Gly Leu Ala Ile Asn Val Ala Val Asn Ile Val Glu Cys
450 455 460

<210> 7
<211> 1437
<212> DNA
<213> Mycobacterium tuberculosis

<220>
<221> CDS
<222> (1)..(1437)
<223> Sequence is identical to GenBank entry GB:MTU87280 [U87280]
Sequence is identical to nucleotides 163-1599 of GenBank entry GB:MTCY427
[Z70692]
Sequence is identical to nucleotides 93-1529 of GenBank entry GB:AE007073

<400> 7
gtg acg gaa aag acg ccc gac gac gtc ttc aaa ctt gcc aag gac gag 48
Met Thr Glu Lys Thr Pro Asp Asp Val Phe Lys Leu Ala Lys Asp Glu
1 5 10 15
aag gtc gaa tat gtc gac gtc cgg ttc tgt gac ctg cct ggc atc atg 96
Lys Val Glu Tyr Val Asp Val Arg Phe Cys Asp Leu Pro Gly Ile Met
20 25 30
cag cac ttc acg att ccg gct tcg gcc ttt gac aag agc gtg ttt gac 144
Gln His Phe Thr Ile Pro Ala Ser Ala Phe Asp Lys Ser Val Phe Asp
35 40 45
gac ggc ttg gcc ttt gac ggc tcg tcg att cgc ggg ttc cag tcg atc 192
Asp Gly Leu Ala Phe Asp Gly Ser Ser Ile Arg Gly Phe Gln Ser Ile
50 55 60
cac gaa tcc gac atg ttg ctt ctt ccc gat ccc gag acg gcg cgc atc 240
His Glu Ser Asp Met Leu Leu Leu Pro Asp Pro Glu Thr Ala Arg Ile
65 70 75 80
gac ccg ttc cgc gcg gcc aag acg ctg aat atc aac ttc ttt gtg cac 288
Asp Pro Phe Arg Ala Ala Lys Thr Leu Asn Ile Asn Phe Phe Val His
85 90 95
gac ccg ttc acc ctg gag ccg tac tcc cgc gac ccg cgc aac atc gcc 336
Asp Pro Phe Thr Leu Glu Pro Tyr Ser Arg Asp Pro Arg Asn Ile Ala
100 105 110
cgc aag gcc gag aac tac ctg atc agc act ggc atc gcc gac acc gca 384
Arg Lys Ala Glu Asn Tyr Leu Ile Ser Thr Gly Ile Ala Asp Thr Ala
115 120 125
tac ttc ggc gcc gag gcc gag ttc tac att ttc gat tcg gtg agc ttc 432

150372450.041602

Tyr	Phe	Gly	Ala	Glu	Ala	Glu	Phe	Tyr	Ile	Phe	Asp	Ser	Val	Ser	Phe	
130						135					140					
gac	tcg	cgc	gcc	aac	ggc	tcc	ttc	tac	gag	gtg	gac	gcc	atc	tcg	ggg	480
Asp	Ser	Arg	Ala	Asn	Gly	Ser	Phe	Tyr	Glu	Val	Asp	Ala	Ile	Ser	Gly	
145				150					155						160	
tgg	tgg	aac	acc	ggc	gcg	gcg	acc	gag	gcc	gac	ggc	agt	ccc	aac	cgg	528
Trp	Trp	Asn	Thr	Gly	Ala	Ala	Thr	Glu	Ala	Asp	Gly	Ser	Pro	Asn	Arg	
				165				170						175		
ggc	tac	aag	gtc	cgc	cac	aag	ggc	ggg	tat	ttc	cca	gtg	gcc	ccc	aac	576
Gly	Tyr	Lys	Val	Arg	His	Lys	Gly	Gly	Tyr	Phe	Pro	Val	Ala	Pro	Asn	
			180				185						190			
gac	caa	tac	gtc	gac	ctg	cgc	gac	aag	atg	ctg	acc	aac	ctg	atc	aac	624
Asp	Gln	Tyr	Val	Asp	Leu	Arg	Asp	Lys	Met	Leu	Thr	Asn	Leu	Ile	Asn	
		195					200					205				
tcc	ggc	ttc	atc	ctg	gag	aag	ggc	cac	cac	gag	gtg	ggc	agc	ggc	gga	672
Ser	Gly	Phe	Ile	Leu	Glu	Lys	Gly	His	His	Glu	Val	Gly	Ser	Gly	Gly	
	210					215					220					
cag	gcc	gag	atc	aac	tac	cag	ttc	aat	tcg	ctg	ctg	cac	gcc	gcc	gac	720
Gln	Ala	Glu	Ile	Asn	Tyr	Gln	Phe	Asn	Ser	Leu	Leu	His	Ala	Ala	Asp	
225					230					235					240	
gac	atg	cag	ttg	tac	aag	tac	atc	atc	aag	aac	acc	gcc	tgg	cag	aac	768
Asp	Met	Gln	Leu	Tyr	Lys	Tyr	Ile	Ile	Lys	Asn	Thr	Ala	Trp	Gln	Asn	
				245					250					255		
ggc	aaa	acg	gtc	acg	ttc	atg	ccc	aag	ccg	ctg	ttc	ggc	gac	aac	ggg	816
Gly	Lys	Thr	Val	Thr	Phe	Met	Pro	Lys	Pro	Leu	Phe	Gly	Asp	Asn	Gly	
			260				265						270			
tcc	ggc	atg	cac	tgt	cat	cag	tcg	ctg	tgg	aag	gac	ggg	gcc	ccg	ctg	864
Ser	Gly	Met	His	Cys	His	Gln	Ser	Leu	Trp	Lys	Asp	Gly	Ala	Pro	Leu	
		275					280					285				
atg	tac	gac	gag	acg	ggc	tat	gcc	ggc	ctg	tcg	gac	acg	gcc	cgt	cat	912
Met	Tyr	Asp	Glu	Thr	Gly	Tyr	Ala	Gly	Leu	Ser	Asp	Thr	Ala	Arg	His	
		290				295					300					
tac	atc	ggc	ggc	ctg	tta	cac	cac	gcg	ccg	tcg	ctg	ctg	gcc	ttc	acc	960
Tyr	Ile	Gly	Gly	Leu	Leu	His	His	Ala	Pro	Ser	Leu	Leu	Ala	Phe	Thr	
305					310					315					320	
aac	ccg	acg	gtg	aac	tcc	tac	aag	cgg	ctg	gtt	ccc	ggc	tac	gag	gcc	1008
Asn	Pro	Thr	Val	Asn	Ser	Tyr	Lys	Arg	Leu	Val	Pro	Gly	Tyr	Glu	Ala	
				325					330					335		
ccg	atc	aac	ctg	gtc	tat	agc	cag	cgc	aac	cgg	tcg	gca	tgc	gtg	cgc	1056
Pro	Ile	Asn	Leu	Val	Tyr	Ser	Gln	Arg	Asn	Arg	Ser	Ala	Cys	Val	Arg	
			340				345						350			
atc	ccg	atc	acc	ggc	agc	aac	ccg	aag	gcc	aag	cgg	ctg	gag	ttc	cga	1104
Ile	Pro	Ile	Thr	Gly	Ser	Asn	Pro	Lys	Ala	Lys	Arg	Leu	Glu	Phe	Arg	

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355	360	365	
agc ccc gac tcg tcg ggc aac ccg tat ctg gcg ttc tcg gcc atg ctg Ser Pro Asp Ser Ser Gly Asn Pro Tyr Leu Ala Phe Ser Ala Met Leu 370 375 380			1152
atg gca ggc ctg gac ggt atc aag aac aag atc gag ccg cag gcg ccc Met Ala Gly Leu Asp Gly Ile Lys Asn Lys Ile Glu Pro Gln Ala Pro 385 390 395 400			1200
gtc gac aag gat ctc tac gag ctg ccg ccg gaa gag gcc gcg agt atc Val Asp Lys Asp Leu Tyr Glu Leu Pro Pro Glu Glu Ala Ala Ser Ile 405 410 415			1248
ccg cag act ccg acc cag ctg tca gat gtg atc gac cgt ctc gag gcc Pro Gln Thr Pro Thr Gln Leu Ser Asp Val Ile Asp Arg Leu Glu Ala 420 425 430			1296
gac cac gaa tac ctc acc gaa gga ggg gtg ttc aca aac gac ctg atc Asp His Glu Tyr Leu Thr Glu Gly Gly Val Phe Thr Asn Asp Leu Ile 435 440 445			1344
gag acg tgg atc agt ttc aag cgc gaa aac gag atc gag ccg gtc aac Glu Thr Trp Ile Ser Phe Lys Arg Glu Asn Glu Ile Glu Pro Val Asn 450 455 460			1392
atc cgg ccg cat ccc tac gaa ttc gcg ctg tac tac gac gtt taa Ile Arg Pro His Pro Tyr Glu Phe Ala Leu Tyr Tyr Asp Val 465 470 475			1437
<210> 8			
<211> 478			
<212> PRT			
<213> Mycobacterium tuberculosis			
<220>			
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<223> Sequence is identical to SwissProt entry SP:GLN1_MYCTU Sequence is identical to PIR entry PIR:H70775 Sequence is identical to PRF entry PRF:2323405A			
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Lys Val Glu Tyr Val Asp Val Arg Phe Cys Asp Leu Pro Gly Ile Met 20 25 30			
Gln His Phe Thr Ile Pro Ala Ser Ala Phe Asp Lys Ser Val Phe Asp 35 40 45			
Asp Gly Leu Ala Phe Asp Gly Ser Ser Ile Arg Gly Phe Gln Ser Ile			

50

55

60

His Glu Ser Asp Met Leu Leu Leu Pro Asp Pro Glu Thr Ala Arg Ile
65 70 75 80

Asp Pro Phe Arg Ala Ala Lys Thr Leu Asn Ile Asn Phe Phe Val His
85 90 95

Asp Pro Phe Thr Leu Glu Pro Tyr Ser Arg Asp Pro Arg Asn Ile Ala
100 105 110

Arg Lys Ala Glu Asn Tyr Leu Ile Ser Thr Gly Ile Ala Asp Thr Ala
115 120 125

Tyr Phe Gly Ala Glu Ala Glu Phe Tyr Ile Phe Asp Ser Val Ser Phe
130 135 140

Asp Ser Arg Ala Asn Gly Ser Phe Tyr Glu Val Asp Ala Ile Ser Gly
145 150 155 160

Trp Trp Asn Thr Gly Ala Ala Thr Glu Ala Asp Gly Ser Pro Asn Arg
165 170 175

Gly Tyr Lys Val Arg His Lys Gly Gly Tyr Phe Pro Val Ala Pro Asn
180 185 190

Asp Gln Tyr Val Asp Leu Arg Asp Lys Met Leu Thr Asn Leu Ile Asn
195 200 205

Ser Gly Phe Ile Leu Glu Lys Gly His His Glu Val Gly Ser Gly Gly
210 215 220

Gln Ala Glu Ile Asn Tyr Gln Phe Asn Ser Leu Leu His Ala Ala Asp
225 230 235 240

Asp Met Gln Leu Tyr Lys Tyr Ile Ile Lys Asn Thr Ala Trp Gln Asn
245 250 255

Gly Lys Thr Val Thr Phe Met Pro Lys Pro Leu Phe Gly Asp Asn Gly
260 265 270

Ser Gly Met His Cys His Gln Ser Leu Trp Lys Asp Gly Ala Pro Leu
275 280 285

<222> (1) .. (1341)

<223> Sequence is identical to complement of nucleotides 4950-6290
of GenBank entry GB:MTCY427 [Z70692]

Sequence is identical to complement of nucleotides 4880-6220
of GenBank entry GB:AE007073

<400> 9

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Met	Asp	Arg	Gln	Lys	Glu	Phe	Val	Leu	Arg	Thr	Leu	Glu	Glu	Arg	Asp		
1				5					10					15			

atc	cgc	ttc	gtc	cgg	ctg	tgg	ttc	aca	gac	gtg	ctc	ggt	ttc	ctc	aag		96
Ile	Arg	Phe	Val	Arg	Leu	Trp	Phe	Thr	Asp	Val	Leu	Gly	Phe	Leu	Lys		
			20					25					30				

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Ser	Val	Ala	Ile	Ala	Pro	Ala	Glu	Leu	Glu	Gly	Ala	Phe	Glu	Glu	Gly		
		35					40					45					

atc	ggc	ttc	gac	gga	tcc	tcg	atc	gag	ggc	ttt	gcg	cgg	gtc	tcg	gaa		192
Ile	Gly	Phe	Asp	Gly	Ser	Ser	Ile	Glu	Gly	Phe	Ala	Arg	Val	Ser	Glu		
	50					55					60						

tcc	gat	acg	gtg	gcg	cac	ccg	gac	ccg	tcg	acc	ttc	cag	gtg	ctg	ccc		240
Ser	Asp	Thr	Val	Ala	His	Pro	Asp	Pro	Ser	Thr	Phe	Gln	Val	Leu	Pro		
65					70					75				80			

tgg	gcc	acc	agt	tcc	ggc	cac	cac	cac	tca	gcg	cgg	atg	ttt	tgc	gac		288
Trp	Ala	Thr	Ser	Ser	Gly	His	His	His	Ser	Ala	Arg	Met	Phe	Cys	Asp		
				85					90					95			

atc	acc	atg	ccg	gac	ggc	tcg	ccg	tcg	tgg	gcg	gac	ccg	cgg	cac	gtg		336
Ile	Thr	Met	Pro	Asp	Gly	Ser	Pro	Ser	Trp	Ala	Asp	Pro	Arg	His	Val		
			100					105					110				

ttg	cgg	cgg	cag	ctg	acg	aag	gcc	ggc	gaa	ctc	ggc	ttc	tcc	tgc	tac		384
Leu	Arg	Arg	Gln	Leu	Thr	Lys	Ala	Gly	Glu	Leu	Gly	Phe	Ser	Cys	Tyr		
		115					120					125					

gtg	cat	ccc	gaa	atc	gag	ttc	ttc	ctg	ctc	aag	ccc	gga	ccc	gag	gac		432
Val	His	Pro	Glu	Ile	Glu	Phe	Phe	Leu	Leu	Lys	Pro	Gly	Pro	Glu	Asp		
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ggg	tcg	gtg	ccc	gtc	ccg	gtc	gac	aac	gcc	ggc	tat	ttc	gac	caa	gcg		480
Gly	Ser	Val	Pro	Val	Pro	Val	Asp	Asn	Ala	Gly	Tyr	Phe	Asp	Gln	Ala		
145				150						155					160		

gtg	cac	gac	tcc	gcc	ttg	aac	ttt	cgc	cgc	cac	gcg	atc	gat	gcc	ctg		528
Val	His	Asp	Ser	Ala	Leu	Asn	Phe	Arg	Arg	His	Ala	Ile	Asp	Ala	Leu		
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gaa	ttc	atg	ggc	atc	tcg	gtg	gag	ttc	agc	cat	cac	gaa	ggc	gca	ccc		576
Glu	Phe	Met	Gly	Ile	Ser	Val	Glu	Phe	Ser	His	His	Glu	Gly	Ala	Pro		
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ggc	cag	cag	gag	atc	gac	ctg	cgg	ttt	gcc	gac	gct	ctg	tcg	atg	gct		624
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Gly	Gln	Gln	Glu	Ile	Asp	Leu	Arg	Phe	Ala	Asp	Ala	Leu	Ser	Met	Ala		
	195						200					205					
gac	aac	gtg	atg	acc	ttc	cgc	tac	gtc	atc	aaa	gaa	gtc	gcg	ctg	gaa		672
Asp	Asn	Val	Met	Thr	Phe	Arg	Tyr	Val	Ile	Lys	Glu	Val	Ala	Leu	Glu		
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gag	ggc	gcc	cgg	gcg	tcg	ttc	atg	ccc	aag	cca	ttc	ggc	cag	cac	ccg		720
Glu	Gly	Ala	Arg	Ala	Ser	Phe	Met	Pro	Lys	Pro	Phe	Gly	Gln	His	Pro		
225					230				235						240		
ggc	tcg	gcg	atg	cac	acc	cac	atg	agc	ctg	ttc	gag	ggt	gat	gtc	aac		768
Gly	Ser	Ala	Met	His	Thr	His	Met	Ser	Leu	Phe	Glu	Gly	Asp	Val	Asn		
				245				250						255			
gcg	ttc	cac	agc	gct	gat	gat	ccg	ctg	cag	ctg	tcg	gaa	gtg	ggt	aaa		816
Ala	Phe	His	Ser	Ala	Asp	Asp	Pro	Leu	Gln	Leu	Ser	Glu	Val	Gly	Lys		
			260				265						270				
tcg	ttc	atc	gcc	ggg	atc	ctg	gag	cac	gct	tgc	gag	atc	agc	gcg	gtc		864
Ser	Phe	Ile	Ala	Gly	Ile	Leu	Glu	His	Ala	Cys	Glu	Ile	Ser	Ala	Val		
		275				280					285						
aca	aat	cag	tgg	gtc	aac	tct	tac	aag	cgg	ctg	gtg	cag	ggc	ggc	gaa		912
Thr	Asn	Gln	Trp	Val	Asn	Ser	Tyr	Lys	Arg	Leu	Val	Gln	Gly	Gly	Glu		
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Ala	Pro	Thr	Ala	Ala	Ser	Trp	Gly	Ala	Ala	Asn	Arg	Ser	Ala	Leu	Val		
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cgg	gtg	ccg	atg	tac	acg	ccg	cac	aag	acc	tcg	tcg	cgg	cgg	gtc	gaa		1008
Arg	Val	Pro	Met	Tyr	Thr	Pro	His	Lys	Thr	Ser	Ser	Arg	Arg	Val	Glu		
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gta	cgc	agc	cct	gat	tcg	gcg	tgc	aat	ccc	tat	ctg	aca	ttc	gcc	gtg		1056
Val	Arg	Ser	Pro	Asp	Ser	Ala	Cys	Asn	Pro	Tyr	Leu	Thr	Phe	Ala	Val		
			340					345					350				
ctg	ctg	gcc	gcg	gga	ttg	cgg	ggt	gta	gag	aag	ggt	tac	gtg	ctg	ggc		1104
Leu	Leu	Ala	Ala	Gly	Leu	Arg	Gly	Val	Glu	Lys	Gly	Tyr	Val	Leu	Gly		
		355				360					365						
ccg	cag	gcc	gag	gac	aac	gta	tgg	gac	ctc	aca	ccc	gag	gaa	cgc	cga		1152
Pro	Gln	Ala	Glu	Asp	Asn	Val	Trp	Asp	Leu	Thr	Pro	Glu	Glu	Arg	Arg		
	370					375					380						
gcg	atg	ggg	tac	cga	gaa	ttg	ccg	tcc	agt	ttg	gat	agt	gcg	ctg	cgc		1200
Ala	Met	Gly	Tyr	Arg	Glu	Leu	Pro	Ser	Ser	Leu	Asp	Ser	Ala	Leu	Arg		
385					390					395					400		
gcc	atg	gag	gcc	tcc	gaa	ctc	gtc	gcg	gag	gcc	ttg	ggg	gag	cac	gtt		1248
Ala	Met	Glu	Ala	Ser	Glu	Leu	Val	Ala	Glu	Ala	Leu	Gly	Glu	His	Val		
				405				410						415			
ttt	gac	ttt	ttc	ttg	cgc	aac	aag	cgc	acg	gag	tgg	gcg	aac	tac	cgc		1296
Phe	Asp	Phe	Phe	Leu	Arg	Asn	Lys	Arg	Thr	Glu	Trp	Ala	Asn	Tyr	Arg		

420

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1341

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<211> 446

<212> PRT

<213> Mycobacterium tuberculosis

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 35 40 45

Ile Gly Phe Asp Gly Ser Ser Ile Glu Gly Phe Ala Arg Val Ser Glu
 50 55 60

Ser Asp Thr Val Ala His Pro Asp Pro Ser Thr Phe Gln Val Leu Pro
 65 70 75 80

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 85 90 95

Ile Thr Met Pro Asp Gly Ser Pro Ser Trp Ala Asp Pro Arg His Val
 100 105 110

Leu Arg Arg Gln Leu Thr Lys Ala Gly Glu Leu Gly Phe Ser Cys Tyr
 115 120 125

Val His Pro Glu Ile Glu Phe Phe Leu Leu Lys Pro Gly Pro Glu Asp
 130 135 140

Gly Ser Val Pro Val Pro Val Asp Asn Ala Gly Tyr Phe Asp Gln Ala

145

150

155

160

Val His Asp Ser Ala Leu Asn Phe Arg Arg His Ala Ile Asp Ala Leu
165 170 175

Glu Phe Met Gly Ile Ser Val Glu Phe Ser His His Glu Gly Ala Pro
180 185 190

Gly Gln Gln Glu Ile Asp Leu Arg Phe Ala Asp Ala Leu Ser Met Ala
195 200 205

Asp Asn Val Met Thr Phe Arg Tyr Val Ile Lys Glu Val Ala Leu Glu
210 215 220

Glu Gly Ala Arg Ala Ser Phe Met Pro Lys Pro Phe Gly Gln His Pro
225 230 235 240

Gly Ser Ala Met His Thr His Met Ser Leu Phe Glu Gly Asp Val Asn
245 250 255

Ala Phe His Ser Ala Asp Asp Pro Leu Gln Leu Ser Glu Val Gly Lys
260 265 270

Ser Phe Ile Ala Gly Ile Leu Glu His Ala Cys Glu Ile Ser Ala Val
275 280 285

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290 295 300

Ala Pro Thr Ala Ala Ser Trp Gly Ala Ala Asn Arg Ser Ala Leu Val
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Val Arg Ser Pro Asp Ser Ala Cys Asn Pro Tyr Leu Thr Phe Ala Val
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355 360 365

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370 375 380

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385 390 395 400

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<222> (1)..(1353)
<223> Sequence is identical to nucleotides 4871-6223
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Sequence is identical to nucleotides 7308-8660
of GenBank entry GB:AE007049

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Glu Gly Val Asp Thr Val Ile Gly Thr Val Val Asn Pro Ala Gly Leu
20 25 30
acc cag gcc aag acc gtg ccg ata cgc cgg acc aac aca ttc gcc aat 144
Thr Gln Ala Lys Thr Val Pro Ile Arg Arg Thr Asn Thr Phe Ala Asn
35 40 45
cct ggc ctc ggc gcc agt ccg gtg tgg cat acc ttc tgt atc gac caa 192
Pro Gly Leu Gly Ala Ser Pro Val Trp His Thr Phe Cys Ile Asp Gln
50 55 60
tgc agt att gca ttc acc gca gac atc agt gtg gtc ggc gat caa cgt 240
Cys Ser Ile Ala Phe Thr Ala Asp Ile Ser Val Val Gly Asp Gln Arg
65 70 75 80
ctc cgc atc gat ctg tcc gcc ttg cgc atc atc ggc gac ggg ttg gcg 288
Leu Arg Ile Asp Leu Ser Ala Leu Arg Ile Ile Gly Asp Gly Leu Ala
85 90 95

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Trp Ala Pro Ala Gly Phe Phe Glu Gln Asp Gly Thr Pro Val Pro Ala	
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Cys Ser Arg Gly Thr Leu Ser Arg Ile Glu Ala Ala Leu Ala Asp Ala	
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Gly Ile Asp Ala Val Ile Gly His Glu Val Glu Phe Leu Leu Val Asp	
130 135 140	
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145 150 155 160	
gcc ggg gtg ctc gag cac gag gcg ttc gtc cgc gat gtc aac gcc gcg	528
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165 170 175	
gca acg gca gca ggc atc gct atc gag cag ttc cat ccc gaa tac ggt	576
Ala Thr Ala Ala Gly Ile Ala Ile Glu Gln Phe His Pro Glu Tyr Gly	
180 185 190	
gcc aac caa ttc gag atc tcg tta gcg ccg cag ccg ccg gtc gcg gcc	624
Ala Asn Gln Phe Glu Ile Ser Leu Ala Pro Gln Pro Pro Val Ala Ala	
195 200 205	
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Ala Asp Gln Leu Val Leu Thr Arg Leu Ile Ile Gly Arg Thr Ala Arg	
210 215 220	
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Ile Gly Ser Gly Ala His Gln His Phe Ser Leu Thr Met Ser Glu Gly	
245 250 255	
atg ctg ttc tcc ggt ggg act gga gca gct ggc atg acc tcg gcc ggg	816
Met Leu Phe Ser Gly Gly Thr Gly Ala Ala Gly Met Thr Ser Ala Gly	
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Glu Ala Ala Val Ala Gly Val Leu Arg Gly Leu Pro Asp Ala Gln Gly	
275 280 285	
atc ctg tgc gga tcg atc gtg tcc ggt ctg cga atg cga ccc ggt aac	912
Ile Leu Cys Gly Ser Ile Val Ser Gly Leu Arg Met Arg Pro Gly Asn	
290 295 300	
tgg gcc gga atc tat gca tgc tgg ggt acc gaa aac cgg gaa gcg gcg	960
Trp Ala Gly Ile Tyr Ala Cys Trp Gly Thr Glu Asn Arg Glu Ala Ala	
305 310 315 320	
gtg cga ttc gtc aag ggc ggg gct ggc agc gcg tac ggc ggg aac gtg	1008

209740 "05450" 05450

Val Arg Phe Val Lys Gly Gly Ala Gly Ser Ala Tyr Gly Gly Asn Val	
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Glu Val Lys Val Val Asp Pro Ser Ala Asn Pro Tyr Leu Ala Ser Ala	
340 345 350	
gcg atc ctc gga ctg gca ctc gac gcc atg aag acc aag gcg gtg ttg	1104
Ala Ile Leu Gly Leu Ala Leu Asp Gly Met Lys Thr Lys Ala Val Leu	
355 360 365	
ccg tcg gaa acg acc gta gac ccg aca cag ctg tct gac gtg gat cgt	1152
Pro Ser Glu Thr Thr Val Asp Pro Thr Gln Leu Ser Asp Val Asp Arg	
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Asp Arg Ala Gly Ile Leu Arg Leu Ala Ala Asp Gln Ala Asp Ala Ile	
385 390 395 400	
gct gta ctg gat agt tcg aaa ctg ctt cgg tgc atc ctt gcc gat ccc	1248
Ala Val Leu Asp Ser Ser Lys Leu Leu Arg Cys Ile Leu Gly Asp Pro	
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Val Val Asp Ala Val Val Ala Val Arg Gln Leu Glu His Glu Arg Tyr	
420 425 430	
ggc gac ctc gat cct gcg cag ctg gcc gac aag ttc cgg atg gct tgg	1344
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35

40

45

Pro Gly Leu Gly Ala Ser Pro Val Trp His Thr Phe Cys Ile Asp Gln
50 55 60

Cys Ser Ile Ala Phe Thr Ala Asp Ile Ser Val Val Gly Asp Gln Arg
65 70 75 80

Leu Arg Ile Asp Leu Ser Ala Leu Arg Ile Ile Gly Asp Gly Leu Ala
85 90 95

Trp Ala Pro Ala Gly Phe Phe Glu Gln Asp Gly Thr Pro Val Pro Ala
100 105 110

Cys Ser Arg Gly Thr Leu Ser Arg Ile Glu Ala Ala Leu Ala Asp Ala
115 120 125

Gly Ile Asp Ala Val Ile Gly His Glu Val Glu Phe Leu Leu Val Asp
130 135 140

Ala Asp Gly Gln Arg Leu Pro Ser Thr Leu Trp Ala Gln Tyr Gly Val
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Ala Gly Val Leu Glu His Glu Ala Phe Val Arg Asp Val Asn Ala Ala
165 170 175

Ala Thr Ala Ala Gly Ile Ala Ile Glu Gln Phe His Pro Glu Tyr Gly
180 185 190

Ala Asn Gln Phe Glu Ile Ser Leu Ala Pro Gln Pro Pro Val Ala Ala
195 200 205

Ala Asp Gln Leu Val Leu Thr Arg Leu Ile Ile Gly Arg Thr Ala Arg
210 215 220

Arg His Gly Leu Arg Val Ser Leu Ser Pro Ala Pro Phe Ala Gly Ser
225 230 235 240

Ile Gly Ser Gly Ala His Gln His Phe Ser Leu Thr Met Ser Glu Gly
245 250 255

Met Leu Phe Ser Gly Gly Thr Gly Ala Ala Gly Met Thr Ser Ala Gly
260 265 270

Glu Ala Ala Val Ala Gly Val Leu Arg Gly Leu Pro Asp Ala Gln Gly
275 280 285

Ile Leu Cys Gly Ser Ile Val Ser Gly Leu Arg Met Arg Pro Gly Asn
290 295 300

Trp Ala Gly Ile Tyr Ala Cys Trp Gly Thr Glu Asn Arg Glu Ala Ala
305 310 315 320

Val Arg Phe Val Lys Gly Gly Ala Gly Ser Ala Tyr Gly Gly Asn Val
325 330 335

Glu Val Lys Val Val Asp Pro Ser Ala Asn Pro Tyr Leu Ala Ser Ala
340 345 350

Ala Ile Leu Gly Leu Ala Leu Asp Gly Met Lys Thr Lys Ala Val Leu
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Pro Ser Glu Thr Thr Val Asp Pro Thr Gln Leu Ser Asp Val Asp Arg
370 375 380

Asp Arg Ala Gly Ile Leu Arg Leu Ala Ala Asp Gln Ala Asp Ala Ile
385 390 395 400

Ala Val Leu Asp Ser Ser Lys Leu Leu Arg Cys Ile Leu Gly Asp Pro
405 410 415

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Ser Val
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<212> DNA
<213> Mycobacterium tuberculosis

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<223> Sequence is identical to complement of nucleotides 3104-4477
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of GenBank entry GB:AE007117

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Met Gln Gly Arg Leu Ala Gly Lys Arg Ile Ser Gly Arg His Phe Val	
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Asp Asp Ile Ala Thr Arg Gly Val Glu Cys Cys Ser Tyr Leu Leu Ala	
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gtg gac gtc gac ctg aac acg gtg ccc ggc tat gcg atg gcc agt tgg	240
Val Asp Val Asp Leu Asn Thr Val Pro Gly Tyr Ala Met Ala Ser Trp	
65 70 75 80	
gac acc ggc tac ggc gat atg gtg atg acg ccg gac ttg tcc act ctg	288
Asp Thr Gly Tyr Gly Asp Met Val Met Thr Pro Asp Leu Ser Thr Leu	
85 90 95	
cgg ctg att cct tgg cta ccg gga acg gcg ctg gtg atc gcc gac ctg	336
Arg Leu Ile Pro Trp Leu Pro Gly Thr Ala Leu Val Ile Ala Asp Leu	
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Val Trp Ala Asp Gly Ser Glu Val Ala Val Ser Pro Arg Ser Ile Leu	
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Arg Arg Gln Leu Asp Arg Leu Lys Ala Arg Gly Leu Val Ala Asp Val	
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Trp Ala Ser Gly Tyr Arg Gly Leu Thr Pro Ala Ser Asp Tyr Asn Ile	
165 170 175	
gac tac gcg ata ttg gca tcc tgc cgg atg gag ccg ttg ctg cgc gac	576
Asp Tyr Ala Ile Leu Ala Ser Ser Arg Met Glu Pro Leu Leu Arg Asp	
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195 200 205

Gly Glu Cys Asn Met Gly Gln Gln Glu Ile Gly Phe Arg Tyr Asp Glu
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225 230 235 240

Glu Ile Ala Asp Gln His Gly Lys Ser Leu Thr Phe Met Ala Lys Tyr
245 250 255

Asp Glu Arg Glu Gly Asn Ser Cys His Ile His Val Ser Leu Arg Gly
260 265 270

Thr Asp Gly Ser Ala Val Phe Ala Asp Ser Asn Gly Pro His Gly Met
275 280 285

Ser Ser Met Phe Arg Ser Phe Val Ala Gly Gln Leu Ala Thr Leu Arg
290 295 300

Glu Phe Thr Leu Cys Tyr Ala Pro Thr Ile Asn Ser Tyr Lys Arg Phe
305 310 315 320

Ala Asp Ser Ser Phe Ala Pro Thr Ala Leu Ala Trp Gly Leu Asp Asn
325 330 335

Arg Thr Cys Ala Leu Arg Val Val Gly His Gly Gln Asn Ile Arg Val
340 345 350

Glu Cys Arg Val Pro Gly Gly Asp Val Asn Gln Tyr Leu Ala Val Ala
355 360 365

Ala Leu Ile Ala Gly Gly Leu Tyr Gly Ile Glu Arg Gly Leu Gln Leu
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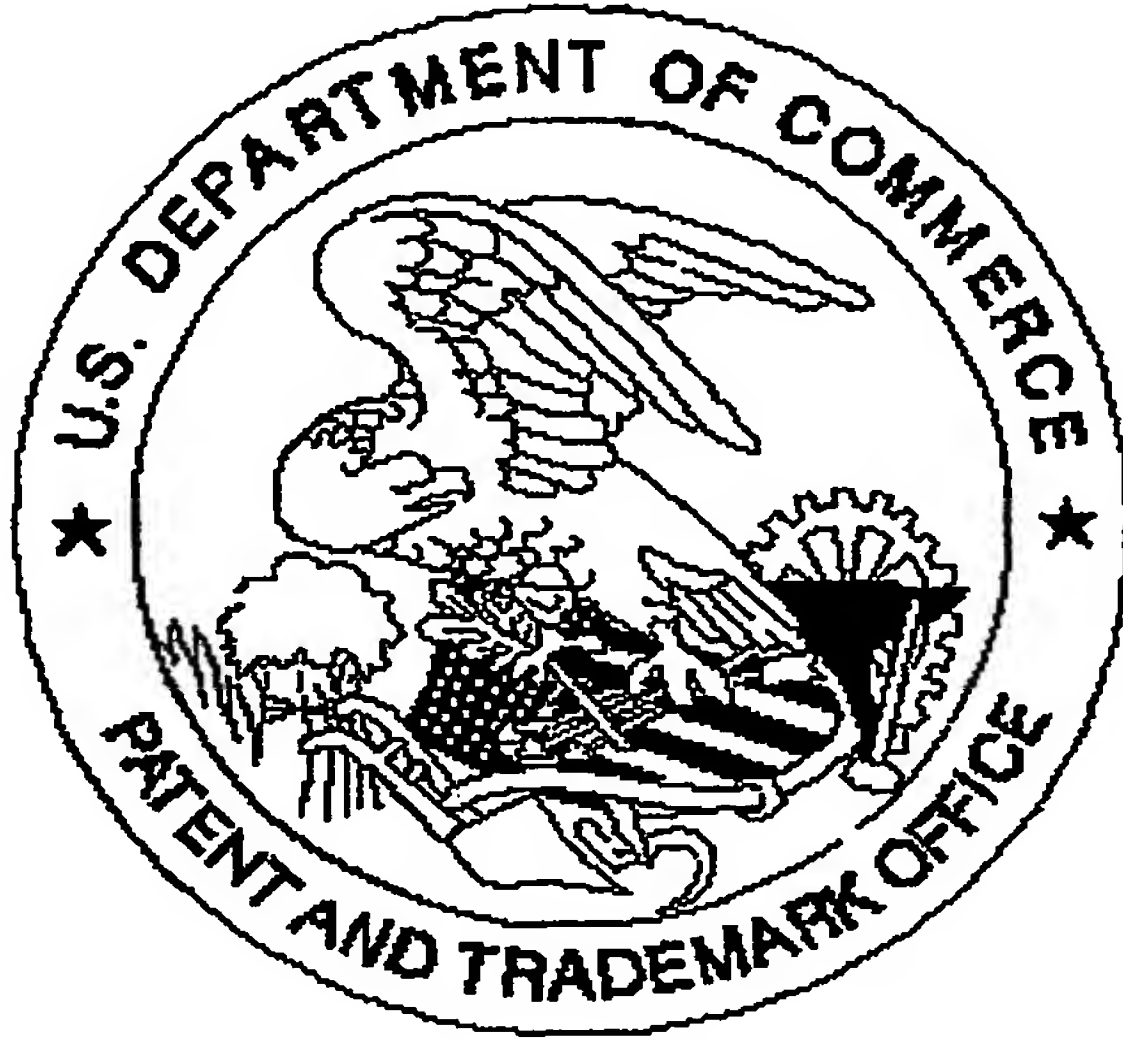
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20240904

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for scanning. (Document title)

☐ Page(s) _____ of _____ were not present
for scanning. (Document title)

☒ **Scanned copy is best available.** Pages numbered 32 to 60 as part of specification are sequence listing.

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